

Synthesis of Some New Substituted 1,3,4-Oxadiazoles and the study of their Biological Activity

Ahmad Kh.Ahmad and Ali O. Obaid
*Chemistry Department, College of Education, University of Mosul
Mosul-Iraq*

(NJC)

(Received on 27/ 9 /2004)

(Accepted for publication on 23/3/2005)

Abstract

Substituted 1,3,4-oxadiazoles are well known as biological active agents. Several compounds of this group were synthesized by using coumarin derivatives as starting material. The structures of the new compounds were established on bases of elemental analysis, physical and spectral data.

-4,3 ,1

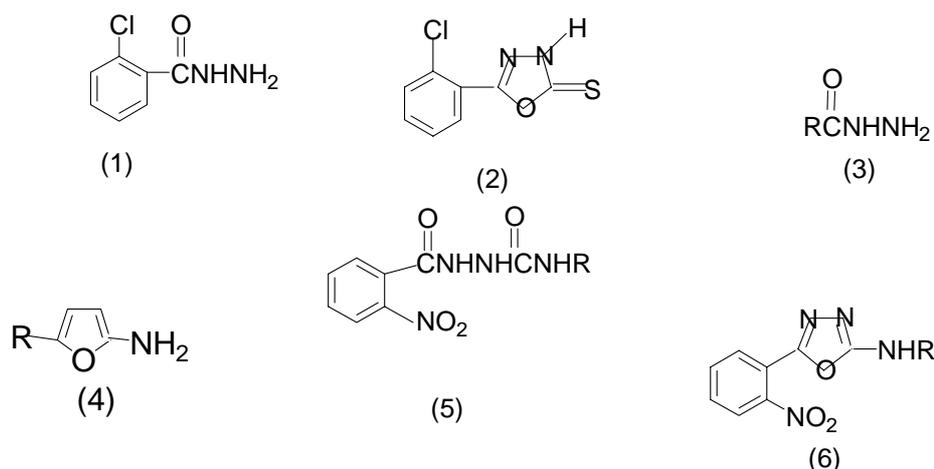
Introduction

1,3,4-oxadiazoles derivatives are known to have biological, medical and industrial applications¹.

Some derivatives show antibacterial² fungicide³ anti-inflammatory and hypotensive action⁴.

The biological activity of these derivatives has drawn the attention of many workers and researchers who attempted to synthesize substituted oxadiazoles. When a mixture of 2-chlorobenzhydrazide (1) and carbon

disulfide in the presence of potassium hydroxide in absolute ethanol was refluxed for 12 hours it offered 3-substituted-5-(2-chlorophenyl)-1,3,4-oxadiazol-2-thione⁵ (2). Other oxadiazole derivatives (4) were obtained by the reaction of hydrazide (3) with cyanogen bromide⁶ and also the reaction of hydrazide with carboxylic acid in the presence of phosphoric acid⁷ or phosphorous oxychloride⁸.



Similarly oxadiazole (6) was obtained by the treatment of (5) with lead oxide⁹, mercury oxide¹⁰ and Dicyclohexylcarbodiimide (Dcc)¹¹. Phosphorous pentoxide was also used for the synthesis of oxadiazole¹².

In this paper the synthesis of some new 1,3,4-oxadiazole is reported.

Experimental

The melting points were measured on Kofler hot stage apparatus and were uncorrected. Elemental analysis measurements of C, H and N were obtained using 1106 CE Carlo-Erba.

The HNMR were recorded on Hitachi Perkin Elemer, R 248, 60 MHz instrument using DMSO-d₆, as a solvent with tetramethylsilane as the internal reference. The IR spectra were recorded on a Pye-Unicam SP 1100 Infra red Spectrophotometer and U.V. spectra were carried out on shimadzu U.V.-Visible recording spectrophotometer.

4-Methyl-7-hydroxy coumarin (7):

This was prepared as mentioned in literature¹³ and yielded 98% m.p. 183-185° (lit. m.p. 185°C).

4-Methyl-7-O- (ethoxy carbonyl methyl) coumarin (8):

This was prepared as mentioned in literature¹⁴ and yielded 97% m.p. 100-102°C (lit. m.p. 102°C).

Ethyl coumarin-3-carboxylate (9):

This was prepared as mentioned in the literature¹⁵ and yielded 77% m.p. 92-90°C (lit. m.p. 94°C)

4-Methyl-7-coumarinloxy methyl hydrazide (10) and coumarin-3-carboxylic acid hydrazide (11)

Method 1¹⁶:

(0.15mol) of ester (8) or (9) and (0.15 mole, 7.26ml) of hydrazine hydrate (99%) in absolute ethanol (100ml) were refluxed for 10 hours. After the reaction mixture was concentrated, when a solid was obtained; it was filtered, washed with cold water and recrystallized from ethanol. The physical and spectral data were given in tables (1 and 2)

Method 2:

(0.05mol.) from ester (8) or (9) with (25ml) hydrazine hydrate (99%) were refluxed for 45min. The product crystallized out on cooling was washed with water, recrystallized from ethanol. See Tables (1 and 2).

1-(4-Methyl-7-coumarinloxy methyl carbonyl)-4-substituted thio-semicarbazide (12-14):

Method 1¹⁷:

An equimolecular quantity of hydrazide (10)(0.002mole, 0.496g) and (0.002mole) substituted isothiocyanate in 40ml absolute ethanol was refluxed for 6 hours. The product was crystallized out on cooling, filtered, recrystallized from ethanol. Tables (3 and 4).

Method 2¹⁷:

Substituted isothiocyanate (0.01mole) and sodium hydroxide (0.01mol, 0.4g as a 2N solution) were added to the solution of compound (10) 0.01 mole in ethanol 20 ml. The mixture was stirred for 24 hours and filtered. The filtrate was acidified with hydrochloric acid. The precipitate was filtered and recrystallized from ethanol-water(50:50).Tables (3and 4).

5-(4-Methyl-7-coumarinloxy methyl)-2-substituted amino-1,3,4-oxa -dizole¹⁸ (15-16).

Dissolve (0.005mole) from any 1-(4-Methyl-7-coumarinyloxy methyl carbonyl)-4-substituted thiosemicarbazide (12-14) in methanol (50ml), then add mercury oxide (0.0055mole, 1.2g.). Refluxed the mixture for 4 hours .The precipitate formed after filtration and evaporation of the solvent is washed with water and recrystallized from ethanol, Tables (5, 6).

5-(4-Methyl-7-coumarinloxy methyl)-2-amino-1, 3, 4-oxadiazole (17) and 5-(3-coumarinloxy)-2-amino-1, 3, 4-oxadizole⁴ (18)

To an ethanolic solution (50ml) of hydrazide (9 or 11)(0.005mole) add cyanogen bromide (0.0055mole, 0.6g). The reaction mixture should be refluxed for 3hours. Cool and neutralize with potassium bicarbonate solution. The solid formed after adding crushed ice is to be filtered and

recrystallized from chloroform. Tables (7,8)

5-(4-Methyl-7-coumarinloxy methyl)-2-substituted-1,3,4,oxadizole (19-27) and 5-(3-Coumarinyl)-2-substituted-1,3,4,oxadizol⁷ (28-33)

An appropriate aromatic carboxylic acid (0.01mole) is added gradually, with stirring for 20 minutes, to a mixture of (0.01mole) of hydrazide (10 or 11) and syrupy phosphoric acid (85%) 10ml at 120°C. The mixture is heated with stirring at this temperature for further 1hour then poured into ice water and left overnight. The precipitate is filtered off, wash with water and with 10% sodium carbonate solution ,recrystallized from ethanol .Tables (9,10)

5-(4-Methyl-7-coumarinyloxy methyl)-1,3,4-oxadizole-2-thiol¹⁹(34)

To a solution containing 100ml of 95% ethanol and 0.01mole of potassium hydroxide, add ((0.01mole, 2.48g.) of hydrazide (10),(0.2 mole, 12ml) of carbon disulfide .Hold the mixture was held at reflux for 4-6 hours(or until most of the hydrogen sulfide is evolved) .After the concentration of the solution to a small volume, the residue is added to ice-water (60g). Acidify with hydrochloric acid (pH =5-6), filter and recystallize the solid from ethanol .m.p. 190°C, 60%. The spectral data is on table (24).

Analysis	C.	H.	N.
Calc.	53.79	3.44	9.65
Found	53.54	3.30	9.52

1-Formyl-2-(4-methyl-7-coumarinloxy methyl carbonyl) hydrazine¹² (35):

A solution of hydrazide (10) (0.025 mole, 6.2 g) in formic acid (98%) (10 ml) is refluxed for 30 minutes. The solvent is evaporated and

the residue is cooled with ice water and recrystallized from methanol. m.p. 178-180°C, 78%.

Analysis	C.	H.	N.
Calc.	56.52	4.34	10.14
Found	56.28	4.29	10.03

2- (4-Methyl -7- Coumarinyloxy methyl) -1,3,4- oxadiazle¹² (36):

To a solution of substituted 1-formyl hydrazine (35) (0.005 mole, 1.38 g) in xylene 100ml, phosphorous pentoxide (0.005 mole, 0.7 g) is added. The mixture is refluxed for 1 hour. The solvent is then evaporated and the residue is recrystallized from methanol. m.p. 98-100°C., 60%

Analysis	C.	H.	N
Calc.	60.46	3.87	10.85
Found	60.08	3.69	10.41

2,5 -bis (substituted)-1,3,4-oxadiazole (37-39):

Phosphoric acid 85% (10 ml) is added to a mixture of (0.0015 mole) hydrazides (10 or 11) and of the esters (0.0015 mole)(8 or 9). The mixture is heated at 120°C for 1 hour Cool and add to ice water and let for 2 hours.

The residue is filtered and washed with water and recrystallized from ethanol, tables (11, 12).

Result and Discussion

The synthesis of new substituted 1,3,4-oxadiazoles may provide additional biologically active agents.

The ester (8) was synthesized by reaction of 4-methyl 7-hydroxy coumarin (7) with ethyl bromo acetate in the presence of anhydrous potassium carbonate, while ethyl coumarin-3-carboxylate (9) was prepared through the reaction of salicylaldehyde with diethyl malonate in the presence of 4-methyl pipyridine. These compounds were identified as mentioned in literatures^{14,15}

Hydrazide (10 or 11) was obtained by refluxing the esters (8 or 9)

with 99% hydrazine hydrate in absolute ethanol. These hydrazides were identified by the appearance of the bands in the infra red spectrum in the following range: bands at 3500-3550 cm^{-1} for (N-H) stretching and also bands for carbonyl groups at (1600-1680 cm^{-1}) which is lower than the carbonyl group of ester due to the presence of resonances effect²⁰.



The ¹HNMR shows that the triplet and quartets bands for ester disappeared and instead of them new bands for (NNH₂) appeared at (3.9 - 4.2) ppm and a broad band at (8.9 - 9.0) ppm for CONH which identified the hydrazide.

Condensation of the hydrazide (10) with suitable alkyl or aryl isothiocyanate provided substituted thiosemicarbazide (12-14) which were identified by IR spectrum as following

stretching bending at (1725-1760 cm^{-1}) for carbonyl group and bands at (3200-3350 cm^{-1}) for N-H stretching as well as bands at (1270-1300 cm^{-1}) for thione groups (C=S). Bands for thiol disappeared and this explains the presence of these compounds in thione form in a solid state⁽²¹⁾. The spectrum of ¹HNMR gave a band between 2-4-2.5 ppm for CSNH and NHCS, also a singlet band appeared at 7.8-8.6 ppm.

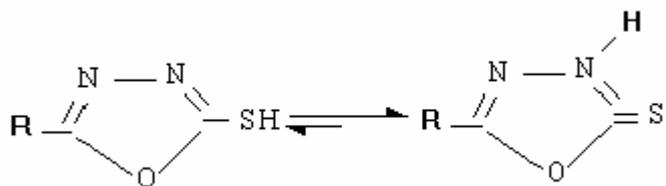
for the CONH proton. Cyclization of compound (12-14) in the presence of mercury oxide in methanol offered the corresponding 5-(3-Methyl-7-coumarinloxy methyl)-2-substituted amino 1,3,4-oxadiazole (15-16). These compounds were identified by spectral data; the spectrum of IR shows the following absorbing bands, at (3150-3380 cm^{-1}) for N-H stretching banding, symmetrical stretching bands at (1230-1260 cm^{-1}) for C-O-C groups and absorption bands at (1610-1640 cm^{-1}) for C=N stretching band.

Also the spectrum of $^1\text{HNMR}$ gave results conforming to expectation, as in table (6). Refluxing hydrazide (10-11) with cyanogen bromide in ethanolic solution gives 5-(4-Methyl-7-coumarinyloxy methyl)-2-amino-1,3,4-oxadiazole (17) and 5-(3-coumarinyl)-2-amino-1,3,4-oxadiazole (18) which were identified by I.R. spectrum through the disappearance of carbonyl band (C=O) for hydrazide and the appearance of band at (1100 cm^{-1}) for symmetrical group C-O-C. A band at (1650 cm^{-1}) for (C=N) stretching group, and bands at (3450-3500 cm^{-1}) were noticed for stretching banding of N-H group. The $^1\text{HNMR}$ spectrum showed bands at (3.0-3.2 ppm) for the absorption of proton NH_2 groups

substituted at oxadiazole ring and other bands appeared at the expected positions as shown in table (8)

Reaction of hydrazide (10 or 11) with different carboxylic acid in the presence of phosphoric acid (85%) at 120°C ; offered 5-(4-Methyl-7-coumarinloxy methyl)-2-substituted-1,3,4-oxadiazole (19-27) and 5-(3-coumarinyl)-2-substituted-1,3,4-oxadiazoles (28-33) which were identified by I.R. spectrum which showed bands at (1090-1180 cm^{-1}) for stretching band of (C-O-C) group. Other bands between (1600-1650 cm^{-1}) for stretching (C=N) group were also noticed. $^1\text{HNMR}$ spectrum showed results as expected. See table (9)

In addition refluxing the hydrazide (10) with carbon disulfide in alcoholic potassium hydroxide gave 5-(4-Methyl-7-coumarinloxy methyl)-1,3,4-oxadiazole-2-thiol (34). This compound was identified by the appearance of stretching band at (1060 cm^{-1}) for (C-O-C) group and the disappearance of stretching band for carbonyl group of hydrazide. Absorption band at (1640-1650 cm^{-1}) for (C=N) and absorption band for (C=S) group appeared at (1200 cm^{-1}) and this confirms the presence of resonance form²⁰



1-Formyl-2-(4-methyl-7-coumarinyloxy methyl carbonyl) hydrazide (35) was obtained by refluxing the hydrazide (10) with formic acid. This compound was identified by IR spectrum which showed a band at (1685 cm^{-1}) for stretching banding of carbonyl group

(C=O) for aldehyde and a band at (1730 cm^{-1}) for carbonyl group of hydrazide. Another band appeared at (3400 cm^{-1}) for N-H stretching band. Cyclization of compound (35) with phosphorous pentoxide in xylene liquid led to the formation 2-(4-Methyl-7-coumarinloxy methyl)-1,3,4-oxadiazole

(36) which was identified by IR Spectrum by the disappearance of the stretching band for carbonyl group of aldehyde and carbonyl group of hydrazide as well as (N-H) group. On the other hand a new band for stretching banding for C=N group appeared at (1100 cm^{-1}) for (C-O-C) group stretching banding.

Reaction of hydrazide (10 or 11) with ester (8 or 9) in the presence of phosphoric acid (85%) at 120°C gave 2,5-disubstituted -1,3,4-oxadiazole (37-

39) and the suggested mechanism for these reaction is as follows;

These compounds were identified by spectrum methods: the IR spectrum showed the disappearance of the following stretching bands for carbonyl ester hydrazide as well as the stretching banding of (C=N) group of hydrazide Bands at $(1640-1650\text{ cm}^{-1})$ for the stretching (C=N) group and bands at $(1050-1130\text{ cm}^{-1})$ for symmetrical stretching band of (C-O-C) group also appeared.

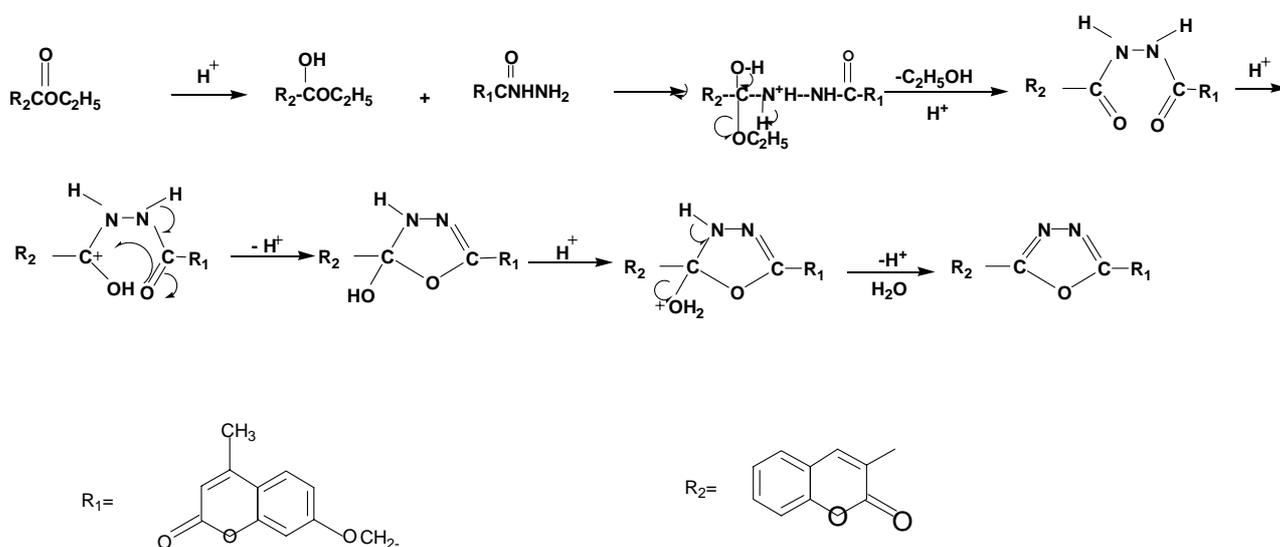


Table (1): physical data for substituted hydrazide



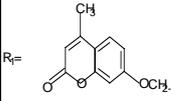
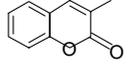
Compd. No.	R	M.p. C°	Yield %	Color	Analysis		
					Calc. C.	Found H.	N.
10		204-206	94	White	58.06	4.83	11.29
11		205-206	85	Yellow	58.82	3.92	13.72
					58.46	4.01	13.58

Table (2): spectral data for substituted hydrazide



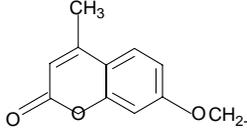
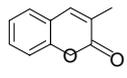
Compd. No.	R	U.V λ_{max} nm EtoH	I.R V cm^{-1}		¹ HNMR δ (ppm) Solv. DMSO-d ₆
			N-H	C=O	
10		248	3500	1680	2.2(s,3H) CH ₃ 4.2(b,2H)NH ₃ 5.1(s,2H)OCH ₂ 6.12-7.5(m,4H)CH, Ar-H 9.0(b,1H)CONH
11		310	3550	1660	4.0(b,2H)NNH ₂ 7.2-7.63(m,4H)Ar-H 8.9(b,1H)CONH 8.72(s,1H)CH

Table (3): Physical data for 1-(4-methyl-7-coumarinloxy methyl carbonyl)-4-substituted thiosemicarbazide

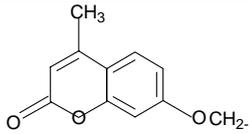
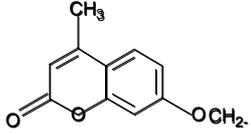
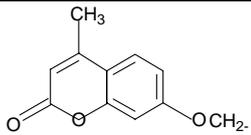
Compd No.	R1	R2	M.P °C	Yield %	color	Analysis		
						Calc./ C.	Found H.	N.
12			178-0	78	White	59.53 59.28	4.43 4.38	10.96 10.79
13		CH3CH2	224-5	60	White	53.73 53.28	5.07 4.97	12.53 12.42
14		CH3	170-2	75	White	52.23 51.98	4.67 4.73	13.08 12.96

Table (4) : Spectral data for 1-(4-methyl-7-coumarinloxy methyl carbonyl)-4-substituted thiosemicarbazide.

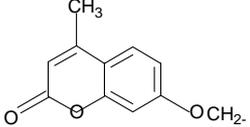
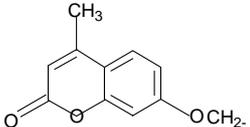
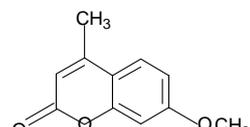
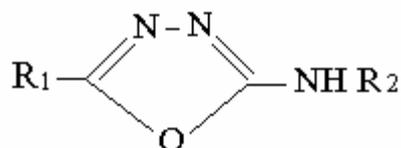
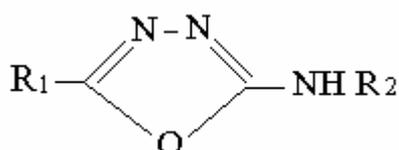
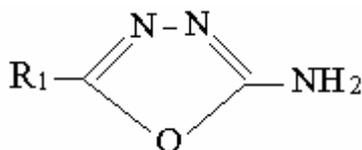
Compd . No.	R1	R2	U.V	I.R Vcm^{-1}		$^1\text{HNMR } \zeta(\text{ppm})$ Solv. DMSO-d6
			λ_{Max} nm EtOH	N-H	C=O C=S	
12			324	3350	1725 1280	2.4(s,3H) CH3 3.0(s,1H) CSNH 3.9(s,1H)NHCS 4.7(s,2H)OCH2 6.0(s,1H)C-H 6.7(m,8H)Ar-H 8.7(b,1H)CONH
13		C2H5	330	3200	1730 1270	1.2(t,3m)CH3 2.3(s,3H)CH3 2.5(s,1H)CSNH 2.8(s,1H)NHCS 3.0(q,2H)CH2 5.0(s,2H)OCH2 6.7-7.5(m,4H)CH,Ar-H 8.7 (b,1H)CONH
14		CH3	333	3300	1760 1300	2.4(s,3H)CH3 2.6(s,3H)CH3 2.7(s,1H)CSNH 2.9(s,1H)NHCS 4.9(s,2H)OCH2 6.1(s,1H)CH 6.7-7.8(m,3H)Ar-H 8.8 (b,1H)CONH

Table (5): Physical data for 5-(4-methyl-7-coumarinloxy methyl) 2-substituted amino-1,3,4-oxadiazole

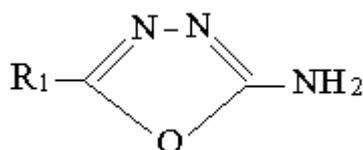
Compd. No.	R1	R2	M.p C°	Yield %	Color	Analysis		
						Calc. C.	Found H.	N.
15			188-0	57	Yellow	65.32	4.29	12.03
						64.97	4.19	11.94
16		CH3 CH2	231-3	63	white	59.80	4.98	13.95
						59.55	5.03	13.82

Table (6) : Spectral data for 5(methyl-7-coumarinyloxy methyl) 2-substituted amino 1,3,4 oxadiazole

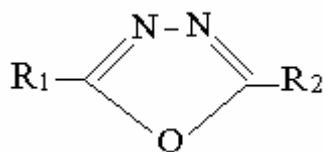
Compd. No.	R1	R2	U.V λ_{max} nm EOH	IR V_{cm}^{-1}		1H NMR ζ (ppm) solv. DMSO-d6
				N-H	C=N C-O-C	
15			302	3380	1640	2.3(S,3H)CH3 2.5(b,1H)NH 4.8(S,2H)OCH2 6.4-8.1(m,9H)Ar-H,CH
16		CH3CH2	348	3150	1610	1.2(t,3H)CH3 2.4(s,3H)CH3 3.3(q,2H)CH2 4.1(b,1H)NH 4.9(s,2H)OCH2 6.6-9.1(m,4H)Ar-H,CH

Table (7): Physical data for 5-substituted 2-amino -1,3,4- oxadiazole

Compd. No.	R1	M.P C°	Yield%	Color	Analysis Calc./Found C. H. N.
17		220-2	73	Pale yellow	57.14 4.02 15.38 56.89 3.98 15.22
18		178-0	50	Brown	57.64 3.05 18.34 57.41 3.00 18.24

Table (8) : Spectral data for 5-substituted 2-amino 1,3,4-oxadiazole

Compd. No.	R	U.V λ_{Max} nm EtOH	I.R Vcm^{-1} N-H,C=H C-O-C	¹ HNMR δ (ppm) Solv. DMSO-d ₆
17		259	3500 1650 1100	2.3(s,3H)CH3 3.0(b,2H)NH2 4.8(s,2H)OCH2 6.1(s,1H)CH 6.7-8.0(m,3H)Ar-H
18		305	3450 1650 1100	

Table (9) : Physical data for 2,5 disubstituted 1,3,4- oxadiazole

Compd. No.	R1	R2	M.P °C	Yield %	color	Analysis		
						Calc./ C.	Found H.	N.
19			188-0	70	White	68.26	4.19	8.38
						67.26	4.23	8.24
20			173-5	2	White	61.87	3.52	7.59
						61.59	3.60	7.43
21			180-2	91	Pale yellow	65.32	4.29	12.03
						64.99	4.32	11.88
22			170-1	85	Pale yellow	53.77	2.83	13.20
						53.28	2.95	13.08
23			218-0	83	White	60.15	3.43	11.08
				[59.79	3.35	10.96
24			193-5	60	White	60.15	3.43	11.08
						59.88	3.38	10.08
25			225-7	53	Yellow	54.81	3.58	9.13
						54.33	3.65	9.08
26			177-9	94	White	64.58	4.03	6.27
						64.21	3.98	6.18
27			140-2	85	White	65.67	3.48	6.96
						65.34	3.41	6.79
28			198-0	62	Yellow	70.34	3.44	9.65
						69.98	3.35	9.49
					Pale brown	62.86	2.77	8.26

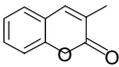
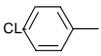
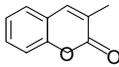
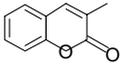
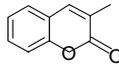
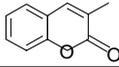
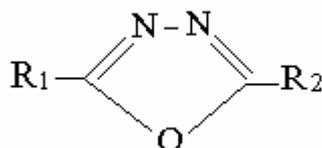
29			190-2	59		62.55	2.70	8.59
			207-9			66.88	3.60	13.77
30			207-9	87	Brown	66.64	3.55	13.70
						43.43	1.50	8.44
31		CL_3C--	210-2	62	Brown	43.21	1.42	8.33
						54.85	2.66	10.66
32		$CLCH_2--$	218-0	75	Pale Yellow	54.54	2.71	10.51
						59.25	3.70	17.28
33		H_2N-CH_2--	217-9	80	Pale yellow	58.89	3.64	17.17

Table (10) : Spectral data for 2,5-disubstituted oxadiazole

Compd No.	R1	R2	U.V λ_{\max} nm EtoH	IR Vcm^{-1} N-H C=N C=S C-O-C	$^1\text{HNMR}$ δ (ppm) Solv.DMSO-d6
19			350	____,1650 ____,1100	2.2(s,3H)CH3 4.4(s,2H)OCH2 6.0(s,1H)CH 6.0-7.6(m,8H)Ar-H
20			348	____,1640 ____,1090	2.4(s,3H)CH3 5.1(s,2H)OCH2 6.1(s,1H)CH 6.6-7.8(m,7H) Ar-H
21			254	3350,1660 1095	4.1(b,2H)NH2 2.2(s,3H)CH3 4.7(s,2H)OCH2 6.2(s,1H)CH 6.5-7.6(m,7H)Ar-H
22			261	____,1640 ____,1105	2.2(s,3H)CH3 4.6(s,2H)OCH2 5.9(s,1H)CH 6.6-8.5(m,6H)Ar-H
23			349	____,1620 ____,1120	2.2(s,3H)CH3 4.6(s,2H)OCH2 6.0(s,1H)CH 6.6-8.4(m,7H)Ar-H
24			346	____,1620 ____,1120	2.2(s,3H)CH3 4.6(s,2H)OCH2 6.0(s,1H)CH 6.6-8.4(m,7H)Ar-H

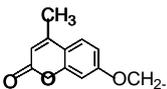
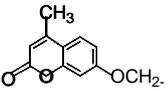
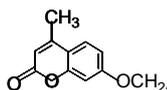
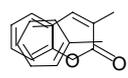
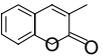
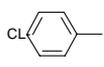
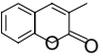
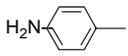
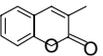
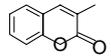
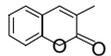
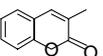
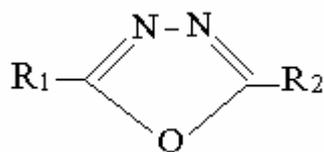
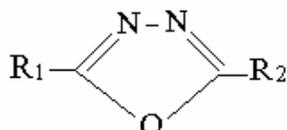
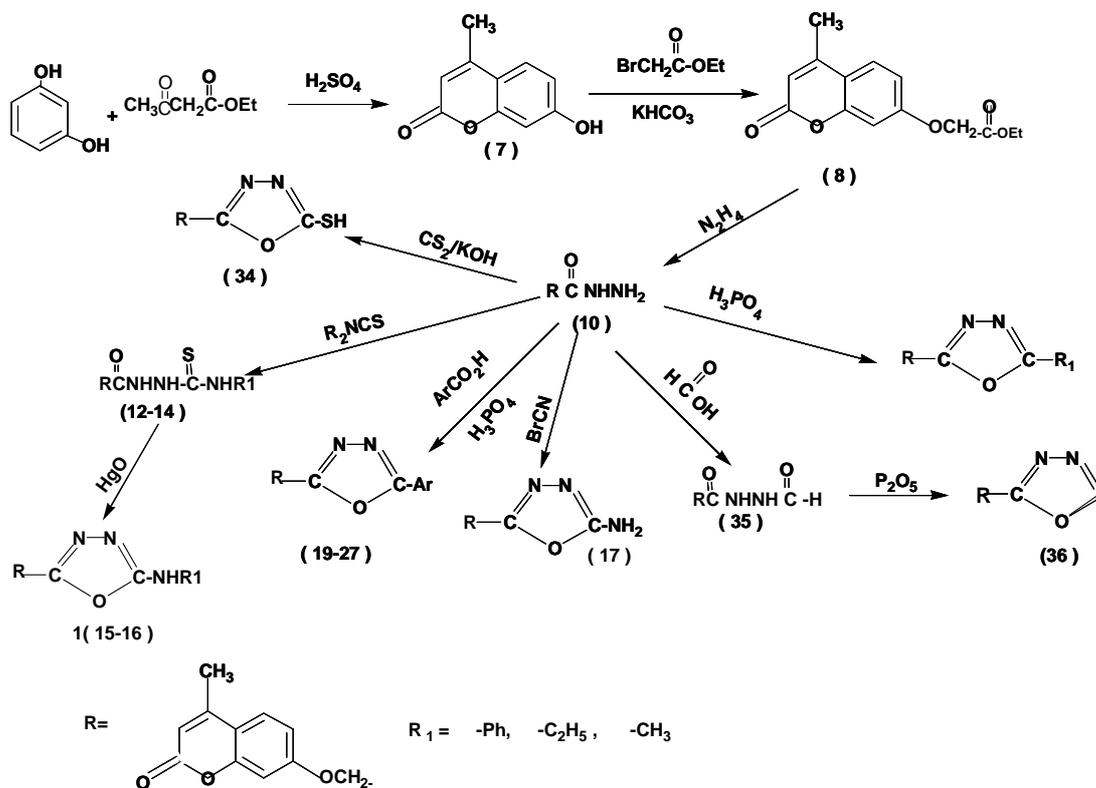
25		CL CH ₂ -	327	— —,1650 —,1110
26		R ₁ --	329	—,1630 —,1120
27			265 245	—,1640 —,1120
28			248	—,1640 —,1110
29			296	—,1640 —,1100
30		C L ₃ C--	310	3300,1620 —,1090
31		CLCH ₂ --	346	— —,1600 —,1150
32		H ₂ N-CH ₂ --	309	—,1610 —,1180
33				3250,1635 ---,1110

Table (11) :physical data for 2,5-bis(substituted)-1,3,4-oxadiazole

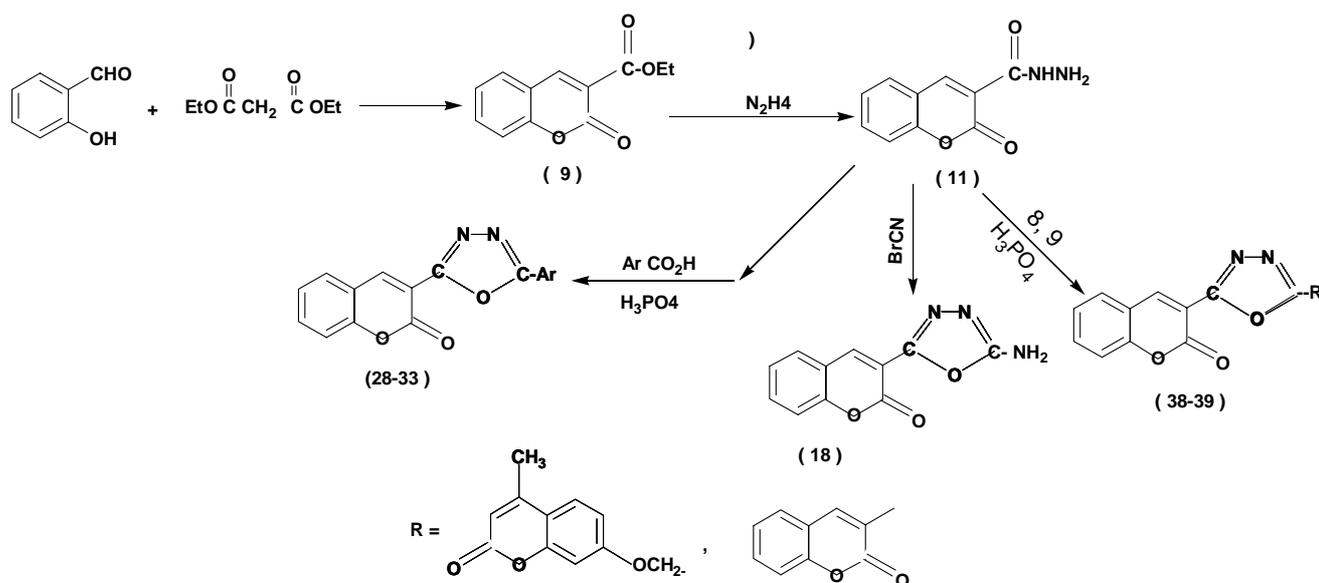
Compd. NO.	R1	R2	M.P. °C	Yield %	color	Analysis		
						Calc/Found		
						C.	H.	N.
37			176-8	91	White	64.57 64.23	4.03 3.95	6.27 6.08
38			163-5	86	Yellow	67.03 66.89	2.79 2.60	7.82 7.63
39			140-2	83	white	65.67 65.34	3.48 3.37	6.96 6.80

Table (12) :Spectral data for 2,5-bis(substituted)-1,3,4-oxadiazole

Compd.No.	R1	R2	U.V λmax nm EtoH	I.R V CM ⁻¹	
				C=N	C-OC
37			330	1650	1050
38			307	1640	1130
39			304	1640	1120



Schem (1) ;Synthesis of oxadiazol derivatives drive from 4-methyl 7-hydroxy coumarine



Schem (2) ; Synthesis oxadiazol derivatives derive from Coumarin-2-Carboxylate

The biological Methods:

1-Bacteria;

The bacteria species used were Gram positive and Gram negative bacteria as listed in Table (13). All strains were obtained from the Department of Biology, College of Science, Mosul University. They were grown up to the stationary phase in a nutrient bath at 37°C and a sample of 0.5 ml of each bacteria was spread over a surface of a nutrient agar plate²².

Sensitivity Test; (Disc Diffusion Method):

Discs of filter paper (6mm diameter) were sterilized at 140°C for 1 hour, impregnated with one of different prepared solution compounds, and then dried-DMSO(dimethylsulfoxide) was used as a solvent for compounds (22, 23, 26, 35). The same solvent was used for antibiotics cephalexine (keflex) for treating pseudomonas aeruginosa, salmonella typhimurium. Cefotaxime (claforan) for Escherichia Coli, staphylococcus aureus and chloramphenicol for Bacillus subtilis were used for comparison.

We also used paper disk of DMSO as control²³. The inoculated plates were incubated at 37° for 14-16 hours, and the inhibition zones (mm) were measured prescott²⁴. In all experiment, the mean of each triplicate was measured²⁵.

Results and Discussion

The antibacterial activities of compounds (22, 23, 31, 34) were evaluated using various species of bacteria:

Staphylococcus aureus, Bacillus subtilis, Pseudomonas aeruginosa, salmonella typhimurium and

Escherichia coli. The results showed that these compounds were active in inhibiting the growth of nearly all micro-organisms and showed that the antibacterial effect on staphylococcus aureus, Bacillus subtilis, Pseudomonas aeruginosa and salmonella typhimurium was higher than that on Escherichia coli as indicated from the diameter of inhibition zone. Table(13).

Blank discs DMSO did not show any activity. The activities of the tested compounds against the studied bacteria were compared with the standard antibiotics. The results showed that some of the compounds had an inhibition zone more than the studied antibiotics, whereas the others showed less effect. It was observed that the compounds (22, 23, 34) had more inhibition zones for Staphylococcus aureus, Bacillus subtilis and Salmonella typhimurium than the antibiotics cefotaxim, Chloroamphenicol and Cephalexine respectively, while compound (31) showed less antimicrobial effect against these microorganism bacteria Table (13).

According to the data (Table 31, 32, 33, 36) it was evident that the activity of tested compounds decreased considerably at lower concentration (0.62 mg /disk). On increasing the Concentration up to (10 mg /disk) a large inhibition zone was observed for these compounds against Pseudomonas aeruginosa.

Table (13): The activities of compounds against bacteria.

Compound	Test Organisms					
	Staph.aures	Bacillis sub	Psedo.ueru	Sal.typh	E.coli	
22	control	S	S+2	MS	S	MS
23		S	S	MS	S	MS
31		MS	S+1	MS	MS	MS
34		S	S	MS	S	MS
Cefotaxime		9				17
Cephalexine				16	10	
Chloramphenicol			8			

Activity of compound (22, 23, 31, 34) against bacteria.

Test Bacteria	Conc. Of 22 mg / disk					Conc. Of 23 mg / disk					Conc. Of 31 mg /disk					Conc. Of 34 mg/disk					
	0.62	1.25	2.5	5.0	10.0	0.62	1.25	2.5	5.0	10.0	0.62	1.25	2.5	5.0	10.0	0.62	1.25	2.5	5.0	10.0	
Staph. aureus	MS	MS	MS	S	S	MS	MS	S	S	MS	MS	MS	MS	MS	MS	MS	MS	MS	S	S	S
BacillusSub.	S	S	MS	S	S+2	MS	MS	S	S	MS	MS	MS	MS	S	MS	MS	MS	MS	S	S	S
Pseud.aerug.	R	R	MS	MS	MS	R	MS	MS	MS	R	MS	MS	MS	MS	MS	MS	MS	MS	MS	MS	MS
SalTyph.	MS	MS	MS	S	S	MS	MS	MS	S	MS	MS	MS	MS	MS	MS	MS	MS	MS	S	S	S
E.coli	R	R	MS	MS	MS	R	MS	MS	MS	MS	R	MS	MS	MS	MS	R	MS	MS	MS	MS	MS

S: Sensitive (Diameter of inhibition zone 6mm less than control sample)

MS: Middle sensitive (Diameter of inhibition zone between 6-12 mm less than control sample)

R: Resistance (Diameter of inhibition zone 12mm larger than control sample)

*: Micro organism under test.

The number of microbial cell in 1mm³ = 1.0 × 10⁸ /ml.

References

1. A.R. Katritzky and C.W. Reez; comprehensive heterocyclic chemistry, pergamon press Ltd., England : 6, 427 (1984).
2. M.A. salama; F.M.A. Moti ; A.A.G. Ghattas and A.abdullah; *Egypt.J.Chem.*, 1981, **24(1)**, 47.
3. V.K. Mishra and S.C. Bahel; *J. Indian Chem. Soc.*, 1983, **LX**, 867.
4. T. Ramalingam ;A.A. Deshmukh;P.B. sattur ; V.K. sheth and S.R. Naik ; *J.indian Chem. Soc.*, 1981, **LVIII**, 269.
5. J.Hazarika and J.C.S. Katakya; *Indian Journal of Heterocyclic Chemistry*, 1998, **7**, 197.
6. K.larc; P.patel; P.vpadhyay and H.Parekh; *Indian Journal of Chemistry*, 1996, **35B**, 1062
7. Shakir M.Said; Ph.D.Thesis,.Mosul university, mosul Iraq (2000)
8. E.H. El-Tamaty; M.E Abdel-Fattah and I.M. El-Deen; *Indian Journal of chemistry* , 1996, **35B**, 1067.
9. J.D. Brooks ; P.T. Charlton ; P.E. Macey; D.A. Peak and W.F. Short; *J. Chem. Soc.*, 1950, **Part I**, 452.
10. U. Srivastava; R.B. pathak and S.C. Bahel; *J. Indian Chem. Soc.* 1982, **LVIII**, 822.
11. S. Sunder; N.p. Peet and R.J. Barbuch ; *J. Heterocyclic Chem.*, 1981, **18**, 1601.
12. A. Shafiee; E. Naimi; P.Mansobi ; F.P. Foroumadi and M. Shekari ; *J. Heterocyclic Chem.*, 1995, **32**, 1235.
13. A.O.Fitton and R.K. Samally; "Practical heterocyclic chemistry", Academic press, London and new York P. 97 (1968)
14. M.I. Husain ; M.K. Shukla and S.K. Agrawal ; *J. Indian Chem. Soc.*, 1979, **LVI**, 306.
15. H.A. Shah and R.C. Shah; *J. Chem. Soc.*, 1939, **Part I**, 132.
16. M.I. Husain ; M.K. Shukla and S.K. Agrawal ; *J. Indian Chem Soc.*, 1979, **LVI**, 306.
17. B.N. Goswami ; J.C.S. Katakya and J.N. Baruah; *J. Heterocyclic Chem.*, 1984, **21**, 1225.
18. M.I. Husain and M.R. Jamali; *Indian Journal of Chemistry*, 1987, **27B**, 43.
19. R.W. Young and K.H. Wood; *J.Am. Chem.Soc.*, 1955, **77**, 400.
20. R.M. Silverstein ; G.C. Bassler and T.C. Morrill, "Spectrometric identification of organic compounds", 3rd Edn., *John wiley & Sons, Inc.*, New York, 1974, **106**, 100.
21. J.P. Henchart; R. Houssain and B. Lablanche; *J. Heterocyclic Chem.*, 1977, **14**, 615.
22. D.A. Shiley; Preparation of Organic Intermediate , *John wiley and Son Inc., New York*, 1951, 243
23. J. Vandepitte ; K.Engbac; P.Pito and G.Heuk; "Basic Laboratory procedure in clinical bacteriology", world Health organization", Geneva, 78, (1991)
24. L.M. Prescott; J.P. Harley and D.A. Klein ; "Microbiology" 3rd. E.d. Wm.c. Brown Publisher , London , Chicago (1996)
25. L.P. Garrod, H.P. Lambert, D. Grady and P. water worth, Antibiotic and chemotherapy, 5th . ed., Churchill living stone, (1981)