

## Synthesis of 2-[(2-Carboxy-4-nitrophenyl) azo]-4,5-diphenyl imidazole (CNPAI) and its usage as an analytical reagent for the determination of Copper (II) and Ferric (III) ions

Abid Allah M. Ali      Sadiq Jaafer Baqir      Ahmed Ali Alkurymy      Ahmed Saadon  
*College of Education      College of Science      College of Science      College of Science*  
*University of Kufa      University of Babylon*

(NJC)

(Received on 21/5/2007) (Accepted for publication on 22/ 8 /2007)

### Abstract

A study for the determination of  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  using 2-[(2-Carboxy-4-nitrophenyl) azo]-4,5-diphenyl imidazole (CNPAI) is performed, the wave length of maximum absorption for Copper (CNPAI) and ferric (CNPAI) complexes are 529 and 533 nm respectively.

After optimum condition were constructed, Beer's law was obeyed in the range (0.4-11) ppm for copper and (0.3-10) ppm for ferric ions, the linearity ( $R^2$ ) was (0.9996), (0.9995) while the molar absorptivity ( $\epsilon$ ) was  $9.5631 \times 10^4 \text{ L.mol}^{-1}.\text{cm}^{-1}$ ,  $7.4414 \times 10^4 \text{ L.mol}^{-1}.\text{cm}^{-1}$  and Sandell sensitivity was  $6.6 \times 10^{-4} \mu\text{g.cm}^{-2}$ ,  $8.7 \times 10^{-4} \mu\text{g.cm}^{-2}$  for Cu(II) and Fe(III) complex respectively.

The stoichiometry of metals to reagent were 1:1 for copper (CNPAI) and 1:2 for ferric (CNPAI) complexes, Precision and accuracy of the analytical procedure for copper and ferric were R.S.D % (1.02, 1.56)% whereas ( $E_{\text{rel}}$ ,  $R_e$ ) % were (-2, 98) %, (-3, 97) %, detection limit was (0.03, 0.056) ppm, the interferences of ions ( $\text{CrO}_4^{2-}$ ,  $\text{Mn}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Ag}^+$ ) were studied and masked by using appropriate masking agents.

(CNPAI)	(III)	(II)
(533 nm)	(529 nm)	
.	(10-0.3)	(11-0.4)

	0.9995	0.9996	(R <sup>2</sup> )
			<sup>4</sup> 10 x 7.4414 <sup>4</sup> 10 x 9.5631
2:1	1:1		
1.02) R.S.D%			
% (98 2-)	(R <sub>e</sub> )	(E <sub>rel.</sub> )	%(1.56
	(0.056)	(0.030)	%(97 3-)
CrO <sub>4</sub> <sup>2-</sup> , Mn <sup>2+</sup> , Co <sup>2+</sup> , Ni <sup>2+</sup> , Zn <sup>2+</sup> , Cd <sup>2+</sup> , Ag <sup>+</sup>			

## Introduction

Azo-dyes with the heterocyclic diazo-component form coloured complexes with many metal ions in solution<sup>(1)</sup>. Great number of the spectrophotometric methods based on these reactions were developed and used in analytical chemistry<sup>(2)</sup>.

In recent years, a lot of researchers deal with the investigations of the mechanism of azo-dyes adsorption on to the solid supports, such as ion-exchangers, PVC, fabrics, silica gel or cellulose<sup>(3)</sup>. The reactions between the immobilized reagents and metal ions in solution have been widely investigated, because of their potentially use in the design of chemical optical sensors<sup>(4,5)</sup>.

The solid phase derivative spectrophotometry using 1-(2-pyridylazo)-2-naphthol (PAN) in Dowex 50W X<sub>2</sub>-100 resin was used for the quantitative determination of copper<sup>(6)</sup>.

Heterocyclic azo dyes are often react with the metal ions as a tridentate ligands forming coordinative bonds with the heterocyclic moiety, -OH or -COOH group and azo group<sup>(7)</sup>.

Many azo compounds are used to determine some metals such as copper and iron<sup>(8-15)</sup>.

Copper is an element that present naturally in many foods, including vegetables, legumes, nuts, grains and fruits, it is a nutritionally essential metal and is widely distributed in nature<sup>(16)</sup>.

It is an essential element not only for life of mammals but also for plants and lower forms of organism, on the other hand it is used algaeicide and herbicide<sup>(17,18)</sup>.

At low levels copper is an essential element in the diet, whereas at higher concentrations it is toxic, because of its

toxicity and cumulative effect in the body<sup>(19)</sup>.

Copper presents a health problem when it occurs in food or water even in minute amounts, hence there is a great need to develop a simple, sensitive, selective and inexpensive method for the determination of copper in environmental biological and soil samples for continuous monitoring to establish the levels of copper in environmental and biological matrices<sup>(20,21)</sup>.

Iron has also a similar importance nearly like copper a number of sensitive analytical methods are available for the determination of these metals, some of the most commonly used methods are spectrophotometric<sup>(22-24)</sup>.

## Experimental

### Apparatus

A UV-Probe model (UV-1650) spectrophotometer (Schimadzu-Japan) and spectronic-21 model U.V-Visible single beam with 1 cm cells Bausch and Lomb (USA) was used for all absorbance measurements, pH measurements were made with Knick-Digital pH meter (England), Digital Balance, Sartorius, (BP 3015-Germany) and Water bath, Gesellschaft Fur Labortechnik (Germany), FTIR 8400S Schimadzu

(Japan) was used to get I.R. spectrums and CHN elemental analyzer 1108 were used.

### Reagents

All chemicals used were of analytical – grade reagent unless otherwise stated, all solutions were prepared using deionized water.

### Synthesis of the reagent (CNPAI)

2-[(2'-Carboxy-4-nitrophenyl) azo]-4,5-diphenyl imidazole (CNPAI) was prepared<sup>(25)</sup> by dissolving (0.9100g, 5mmol) of 5-nitroanthranilic acid in 30 ml of water and 3 ml of concentrated hydrochloric acid.

The filtered solution was cooled to 0°C, treated with 8 ml of aqueous (1 M) sodium nitrite drop wise, and stirred for 30 min.

A 4,5-diphenyl imidazole (1.1000g, 5mmol) was dissolved in 100 ml of pyridine, and 5 ml of 10% sodium hydroxide and 25 ml of 10% sodium carbonate were added, the diazonium solution prepared above was then added drop wise for coupling, after the mixture had been stirred for an hour at 5°C, it was acidified with dilute hydrochloric acid to pH = 5.

The precipitate was filtered off, and recrystallized twice from hot ethanol, and then dried in the oven at 60°C for several hours, the yield was (54%) (2.2300g) of reddish brown powder

(m.p = 112-114 °c), with the formula  $C_{22}H_{15}N_5O_4$  with the theoretical value C= 63.92%, H= 3.63%, N= 16.94% while the element analysis gave the value of C= 63.58%, H= 3.45% and N= 16.78%.

#### **Standard Copper & Ferric solutions**

Solution of  $Cu^{2+}$  and  $Fe^{3+}$  (100 ppm) were prepared by dissolving (0.0268 g) of  $CuCl_2 \cdot 2H_2O$  and (0.0290g) of  $FeCl_3$  in 100 ml distilled water, working solution were prepared freshly by appropriate dilution of the stock solution.

#### **CNPAI Solution**

A solution of ( $1 \times 10^{-3}$  M) was prepared by dissolving (0.0413g) of pure reagent in 100 ml of absolute ethanol.

#### **General procedure**

Into a 5 ml calibrated flask, transfer (0.5 ml) of sample solution containing not more than 2 ppm and 1.5 ppm of the ion ( $Cu^{2+}$  or  $Fe^{3+}$ ) and (0.5 ml) of  $1 \times 10^{-4}$  M ethanolic reagent (CNPAI) solution dilute to volume with deionized water, mix well and after 10

minutes measure the absorbance at 529 nm for  $Cu^{2+}$  and 533 nm for  $Fe^{3+}$  in a 1 cm cell against a blank solution prepared in a similar way but without the presence of the ion under test.

### **Results & Discussion**

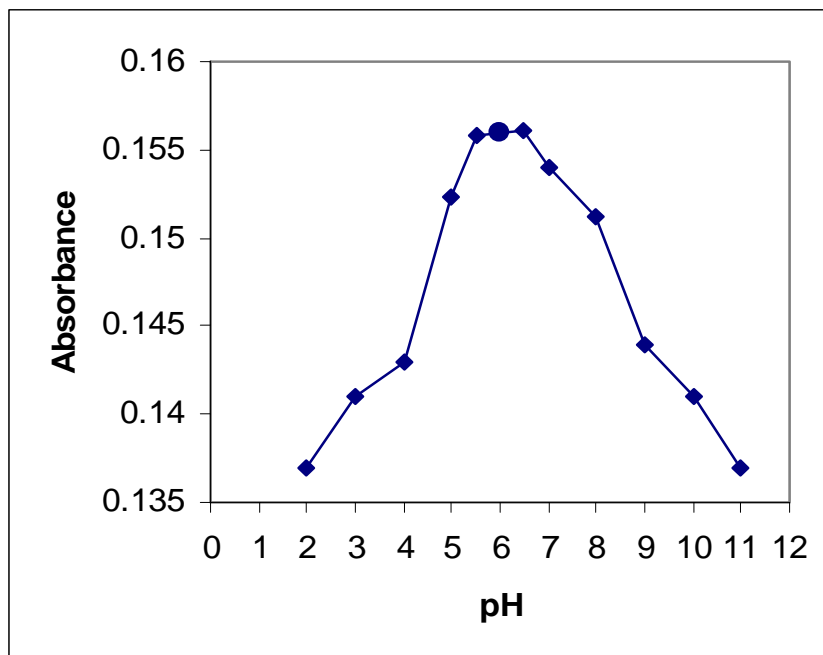
#### **Physical and chemical properties of CNPAI**

The reagent is a reddish brown powder which is sparingly soluble in water. It has a good solubility in ethanol, methanol, acetone, chloroform and ether.

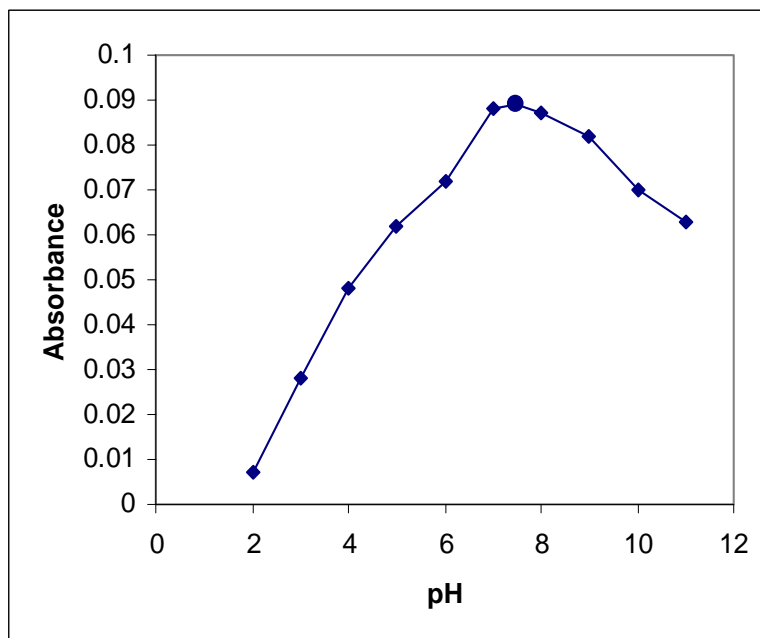
The color of the solution is red in alkaline medium, yellow in weakly and strong acidic solution.

#### **Effect of pH**

The effect of acidity on the absorbance value of the complex was investigated by changing the pH value of the solution and the results are shown in, figures (1&2).



**Fig.(1) Effect of pH on the absorbance of copper complex at room temp.**



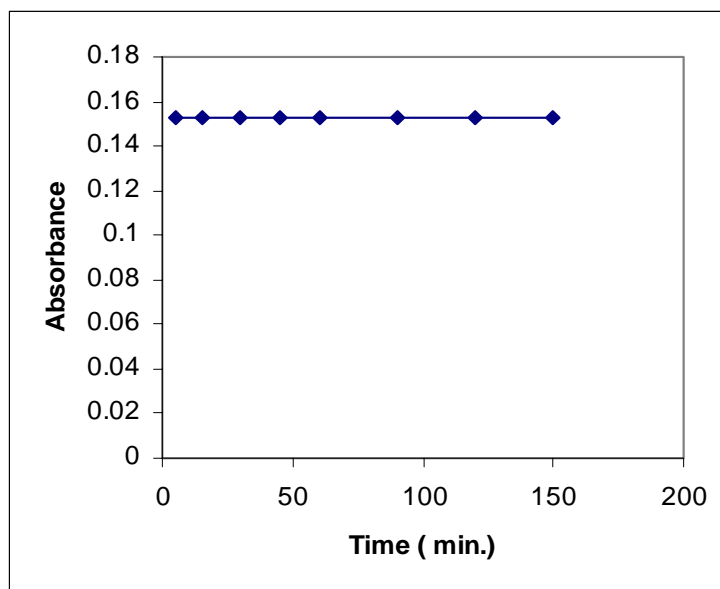
**Fig.(2) Effect of pH on the absorbance of ferric complex at room temp.**

Figures (1&2) demonstrated that the best absorbance of Cu(II) CNPAI system is in the range (5.5-6.5) and pH (6) was adopted as optimum while for Fe(III) CNPAI system is in the range (7-8) and the pH (7.5) was adopted as

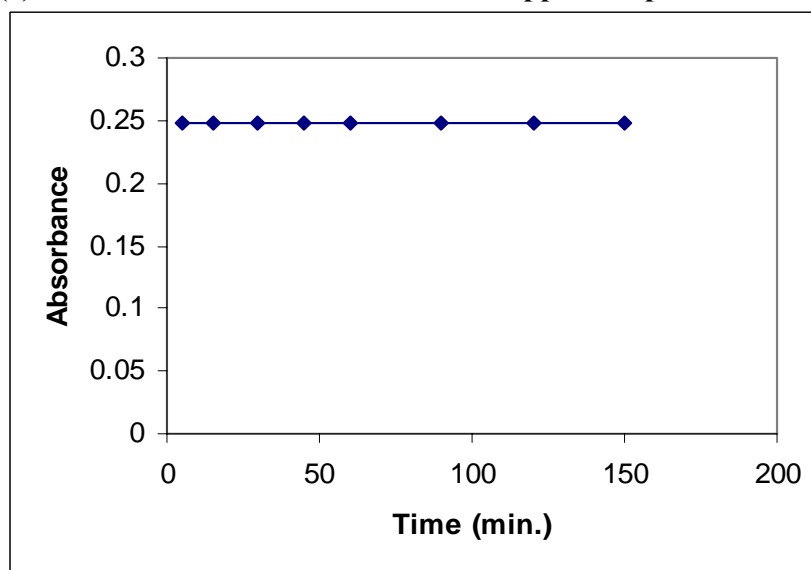
optimum pH.

#### **Stability of the chromogenic system with the time**

Stability of the chromogenic system with the time is shown in figures (3&4).



**Fig.(3) Effect of time on the absorbance of copper complex at room temp.**

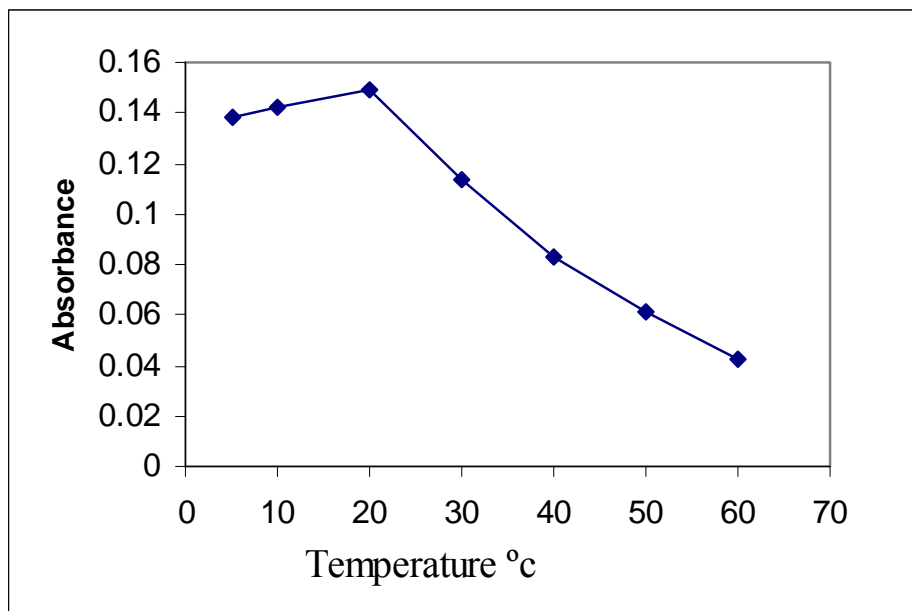


**Fig.(4) Effect of time on the absorbance of ferric complex at room temp.**

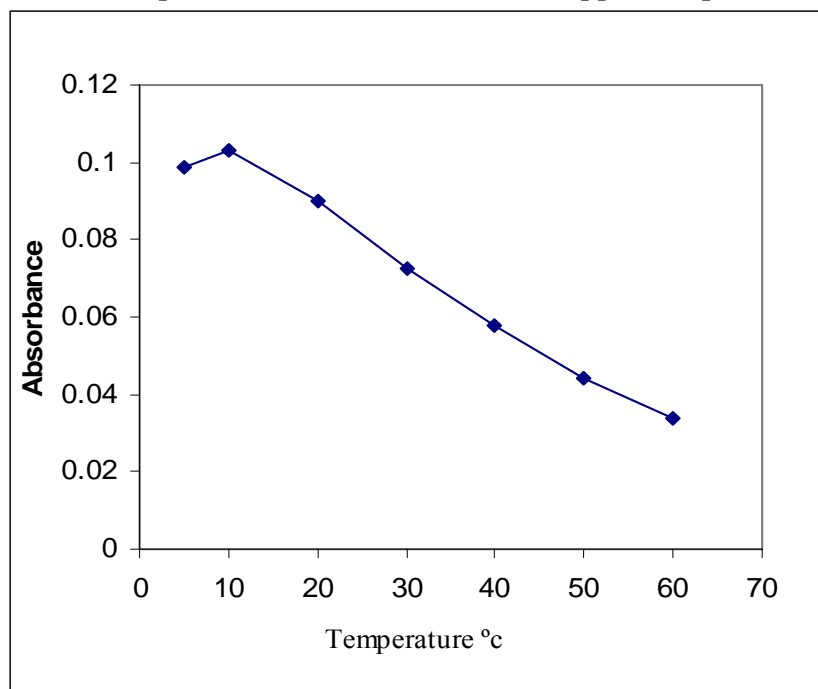
Figs. (3&4) show that the color of the complex system reaches its maximum value of absorbance from (5) min. and remain stable for about (24) hours.

#### **Effect of temperature.**

The effect of temperature on the absorbance of the two complexes; Cu(II)-CNPAI and Fe(III)-CNPAI was studied figures (5&6) show this effect.



**Fig.(5) Effect of temperature on the absorbance of copper complex.**



**Fig.(6) Effect of temperature on the absorbance of ferric complex.**

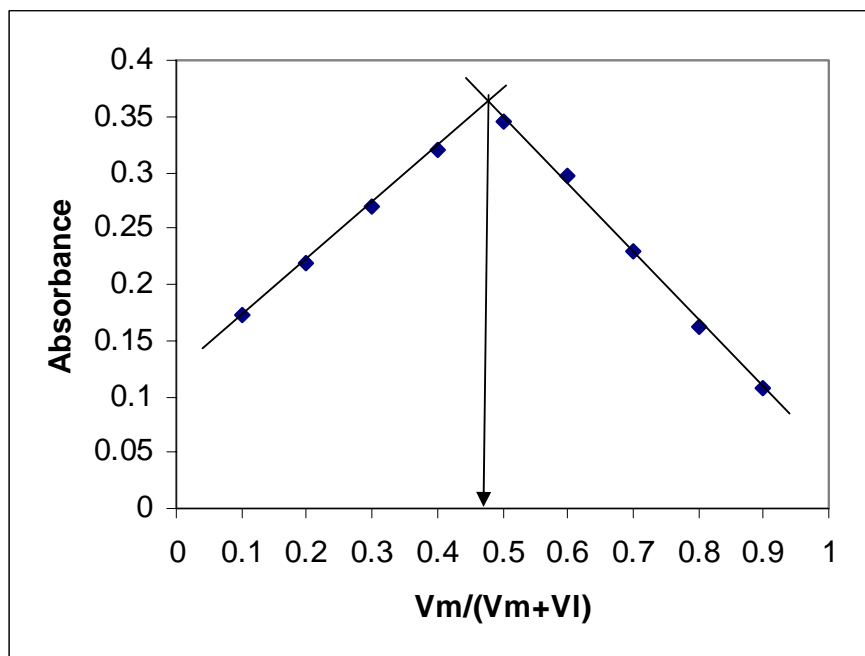
The study was performed at temperatures at the range (10-60) °c, and the maximum absorption was obtained at 20 °c for Cu(II)-CNPAI and 10 °c for Fe(III)-CNPAI

complexes, the decrease in absorbance value may be is due to the dissociation of the complex.

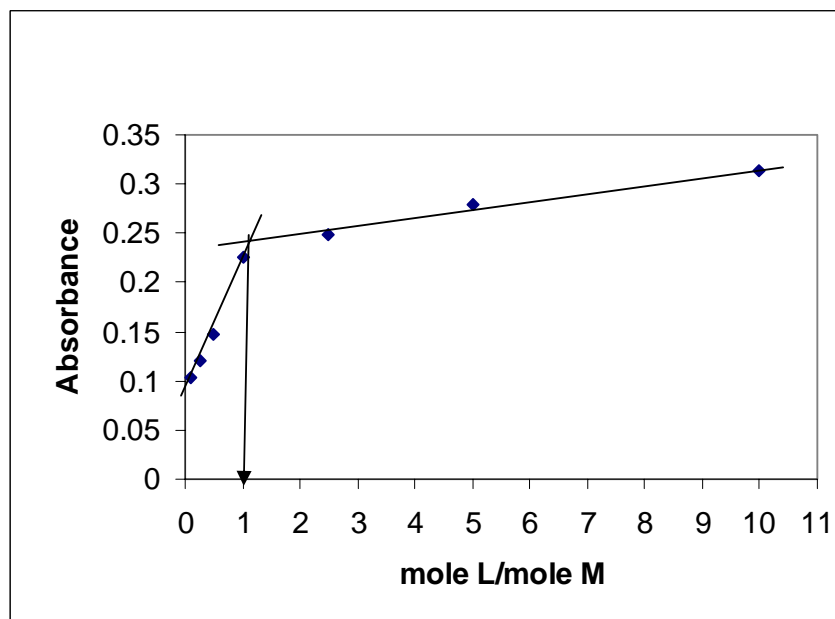
### Composition of the complexes

The composition of the two complexes was determined by Job's<sup>(26)</sup> method of continuous variation and molar ratio<sup>(27)</sup>

methods which were applied to ascertain the stoichiometric composition of each of the complexes as shown in figures (7-10).

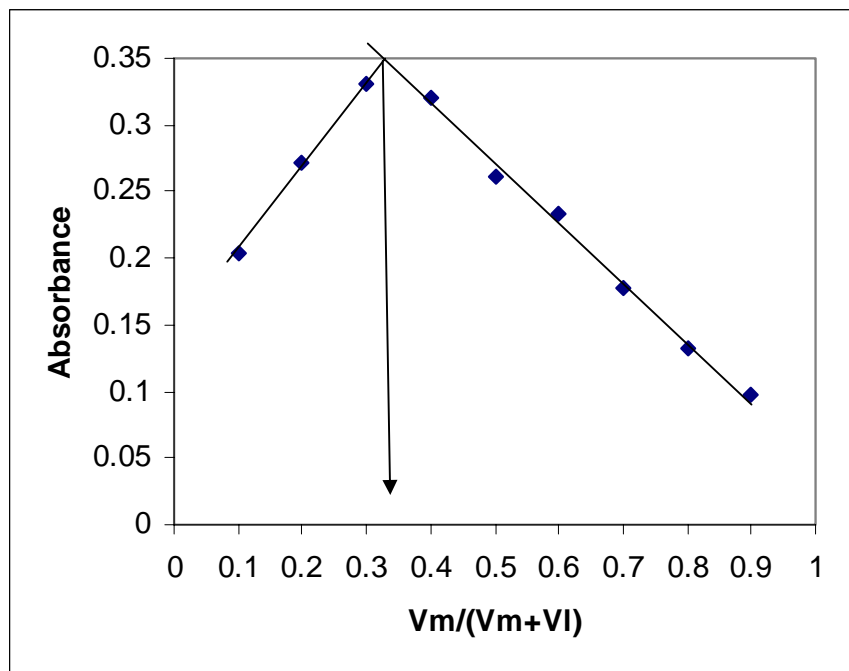


**Fig.(7)** continuous variation method for Cu(II)-CNPAI complex at optimum conditions.

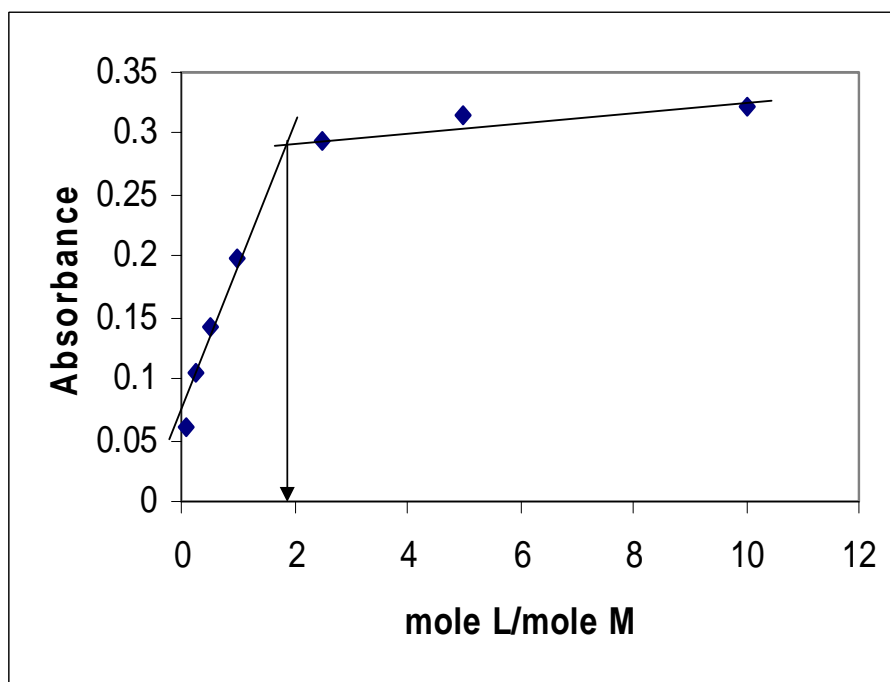


**Fig.(8)** molar ratio method for Cu(II)-CNPAI complex at optimum condition.





**Fig.(9)** continuous variation method for Fe(III)-CNPAI complex at optimum conditions.

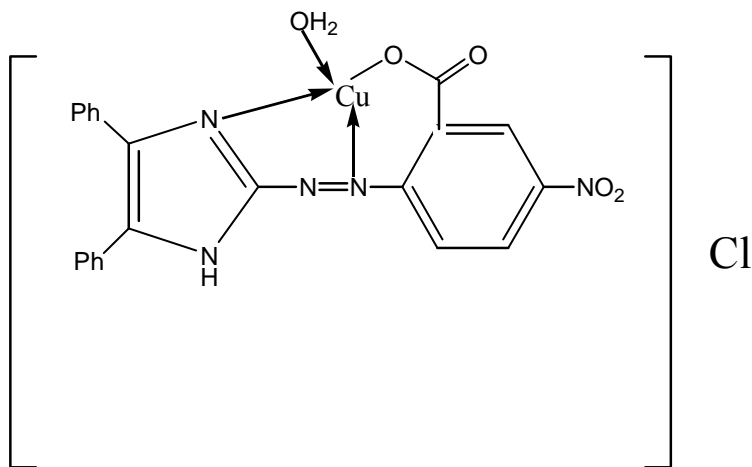


**Fig.(10)** molar ratio method for Fe(III)-CNPAI complex at optimum conditions.

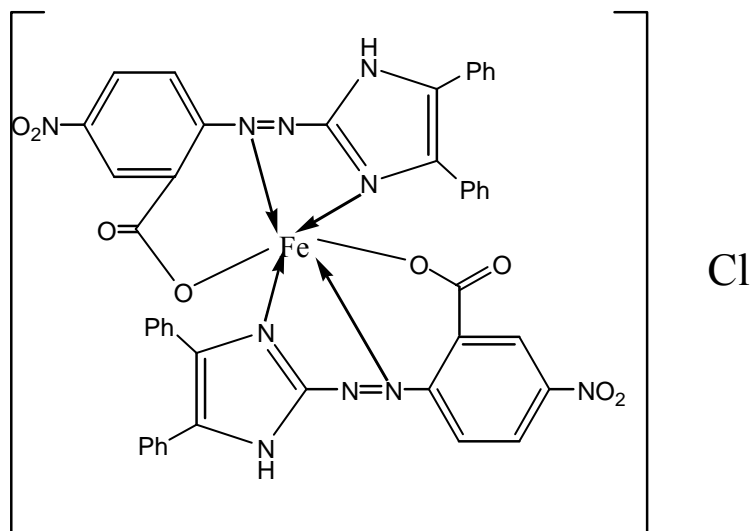
From the results of continuous variation and molar ratio methods show in figures (7-10), the ratio

between copper to CNPAI is (1:1), and between ferric to CNPAI is (1:2) and the stability constant<sup>(28)</sup> ( $K_{sta.}$ ) was

$(4.13 \times 10^7)$   $\text{L} \cdot \text{mol}^{-1}$  and  $2.61 \times 10^8$  (11,12) shows the proposed  
 $\text{L}^2 \cdot \text{mol}^{-2}$ ) for Cu(II)-CNPAI and composition of the two complexes  
 Fe(III)- CNPAI complexes, figures



**Fig. (11) the composition of Cu-CNPIA complex**



**Fig. (12) the composition of Fe-CNPIA complex**

### Calibration Graphs

Calibration graphs for Cu (II) and Fe (III) complexes were obtained by

following the proposed procedure under the optimum conditions.

These are shown in figures (13&14).

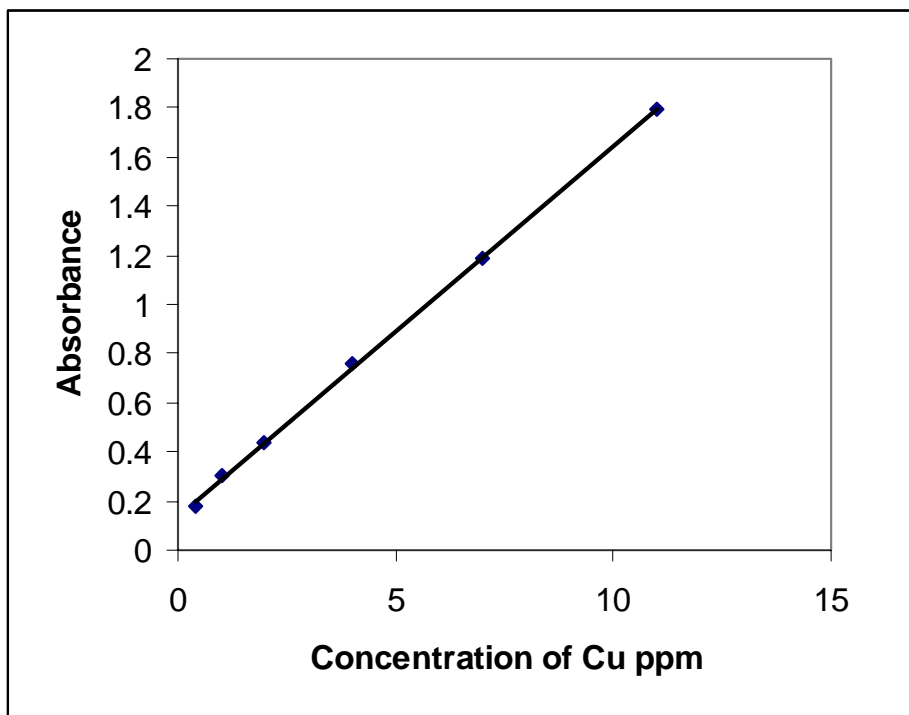


Figure (13) calibration graph of Cu (II) ion.

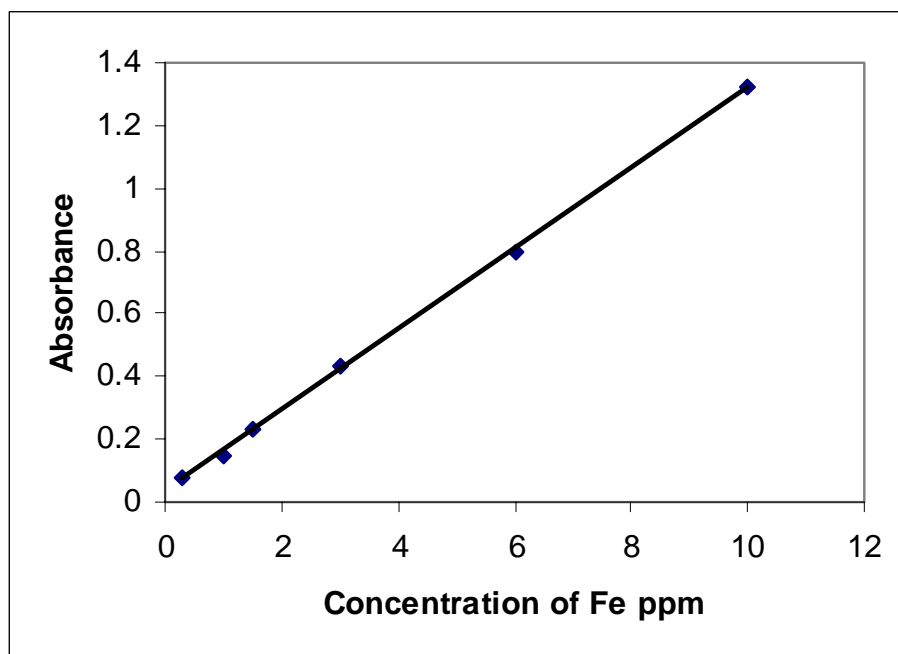


Figure (14) calibration graph of Fe (III) ion.

The results indicated that Beer's law was obeyed over the concentration range (0.4-11) ppm for Cu (II) and (0.3-10) ppm for Fe (III), linearity ( $R^2$ )

for copper was (0.9996) and ferric was (0.9995) respectively.

The molar absorption coefficient and Sandell's sensitivity were calculated to

be  $(9.5631 \times 10^4)$ ,  $(6.6 \times 10^{-4})$  for Cu (II) and  $(7.4414 \times 10^4)$ ,  $(7.6 \times 10^{-4})$  for Fe (III).

The detection limit was (0.03) ppm for Cu (II) and (0.056) ppm for Fe (III).

#### Precision and Accuracy

The precision of the present analytical method was evaluated by determining different concentrations of Cu (II) and Fe (III) each analyzed (5 times), the relative standard deviation R.S.D % was (1.02) and (1.56) respectively indicating that this method is highly precise and reproducible.

The reliability of the method was tested by recovery (Re %) were (-

2,98) for Cu (II) and (-3, 97) for Fe (III).

The precision and accuracy of the method were found to be excellent.

#### Interferences

The selectivity of copper (CNPAI) and ferric (CNPAI) complexes is tested by measuring the absorbance of complex of 2 ppm of Cu (II) and 1.5 ppm Fe (III) at optimum conditions in presence of different foreign ions of  $5 \times 10^{-4}$  M concentration which are able to form complexes with (CNPAI)<sup>(29)</sup>.

The extent of reaction of these ions is shown in table (1)

**Table (1) effect of interference ions**

Interference ions $5 \times 10^{-4}$ M	Interference %	
	Cu <sup>2+</sup> -(CNPAI)	Fe <sup>3+</sup> -(CNPAI)
Cd <sup>2+</sup>	+3.30	+2.84
Co <sup>2+</sup>	+6.45	+5.23
Ni <sup>2+</sup>	+7.72	+2.63
Zn <sup>2+</sup>	+5.49	+3.05
Mn <sup>2+</sup>	+1.22	+4.13
Ag <sup>1+</sup>	+1.43	+0.85
CrO <sub>4</sub> <sup>2-</sup>	-1.52	-2.45
Fe <sup>3+</sup>	+5.25	-----
Cu <sup>2+</sup>	-----	+8.12

The criterion for interference<sup>(30)</sup> was an absorbance value varying by more than 5% from the expected value for copper (II) or iron (III) complex so there is an obvious interference of Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup> and Fe<sup>3+</sup> with Cu-(CNPAI) complex while Fe<sup>3+</sup>-(CNPAI)

complex is effected by Co<sup>2+</sup> and Cu<sup>2+</sup> interference.

#### Effect of masking agents<sup>(31)</sup>

The effect of masking agents was studied to increase the selectivity of complexes, this effect is shown in table (2).

**Table (2) the effect of masking agents**

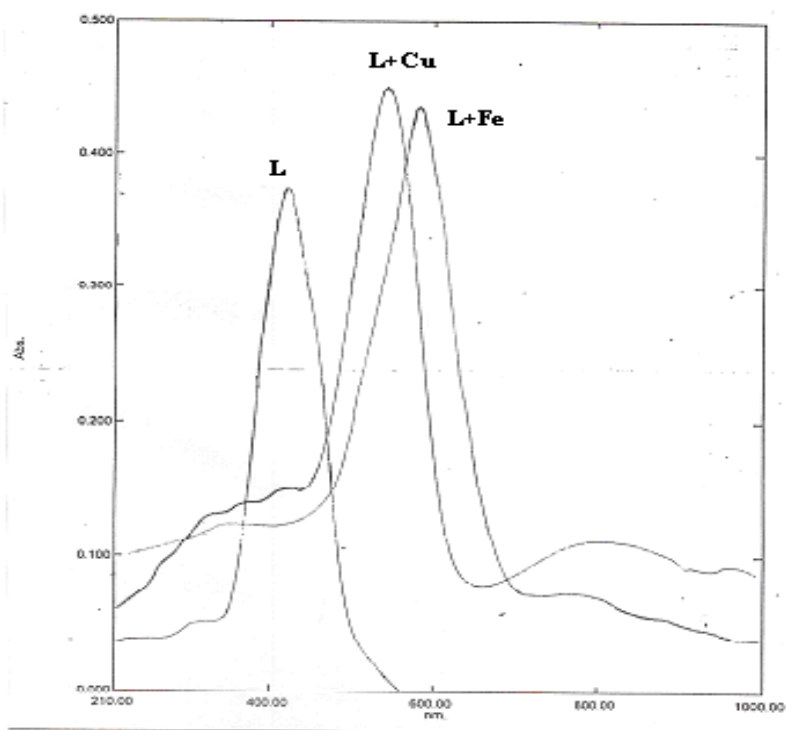
Masking agents $1 \times 10^{-2}$ (M)	Absorbance of $1 \times 10^{-4}$ M	
	Cu (II)-CNPAI	Fe (III) -CNPAI
Complex without any addition	0.153	0.038
Tartaric acid	0.103	0.009
Oxalic acid	0.082	0.015
Citric acid	0.108	0.049
Ascorbic acid	0.075	0.020

The results shown in table (2) indicates that ascorbic acid and oxalic acid is better masking agents for copper complex while tartaric acid and oxalic acid are better for ferric complex,

other masking agents are less effect comparatively.

#### Absorption spectra

The absorption spectra of the two complexes and the ligand are shown in figure (15) under optimum conditions



**fig.(15) absorption spectra of the two complexes and the ligand.**

Absorption spectra show that the  $\lambda$  max of absorption of the reagent (CNPAI) is obvious at 425 nm, Cu (CNPAI) complex at 529 nm and Fe (CNPAI) complex at 533 nm, this new  $\lambda$  max mean red shift in  $\lambda$  max of complexes.

#### **FTIR spectrum of (CNPAI) reagent**

The structure of CNPAI reagent was demonstrated by using FTIR technique using KBr disc, fig.(16) show the FTIR spectrum.

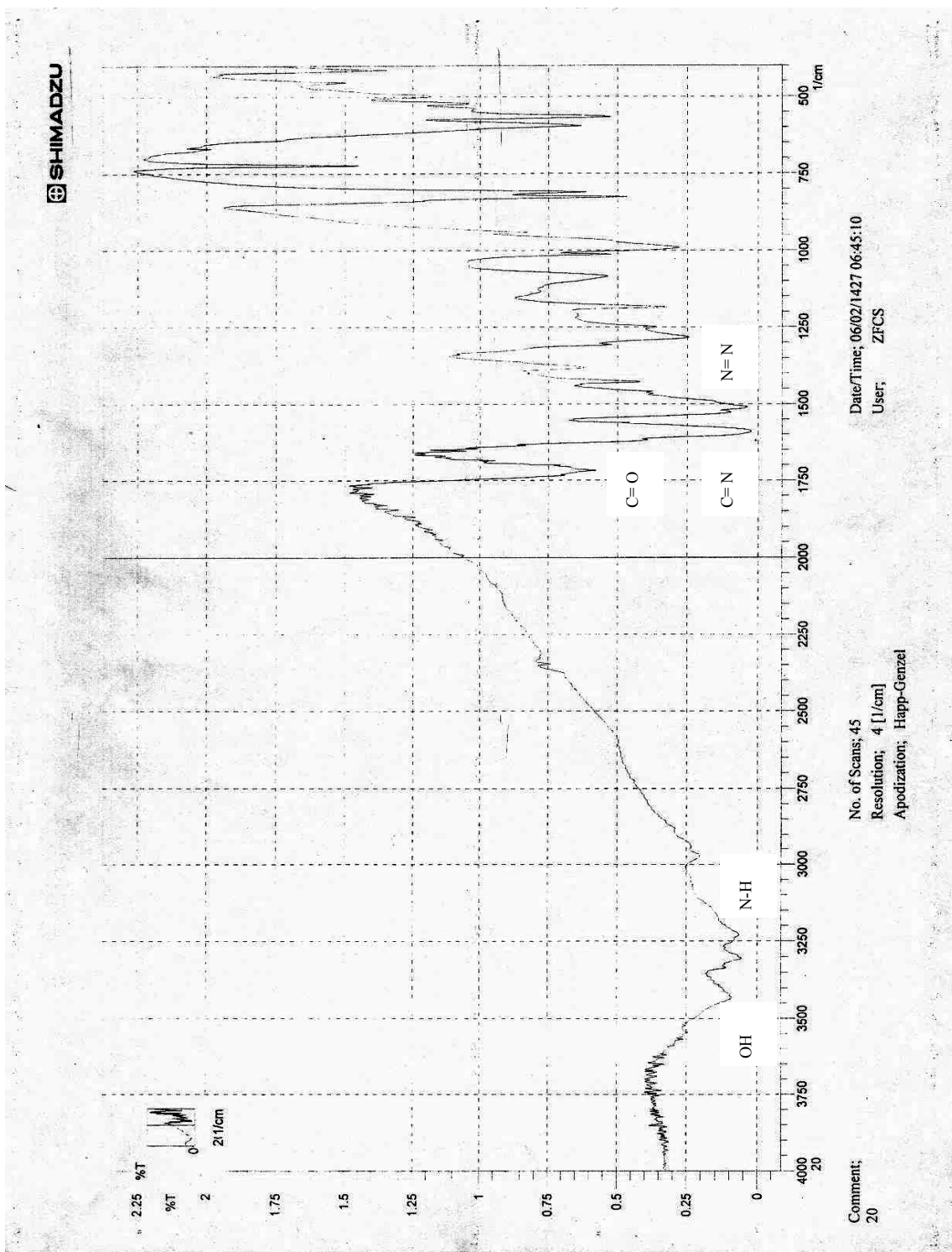


Figure (16) FTIR spectrum of (CNPAI).

The infrared spectrum show in figure (14) give an evidence for the formation of the reagent (CNPAI) where the broad absorption band at (3350-3450)  $\text{cm}^{-1}$  referred to the stretching vibration  $\nu$  (N-H) of the imidazol ring<sup>(32)</sup>

A weak band appeared at (2980)  $\text{cm}^{-1}$  due to  $\nu$  (C-H), the band at (1690)  $\text{cm}^{-1}$  referred to the  $\nu$  (COOH) group, the band at (1590)  $\text{cm}^{-1}$  referred to the  $\nu$  (C=N) stretching of the heterocyclic imidazol ring, at (1500)  $\text{cm}^{-1}$  the stretching vibration  $\nu$  (N=N) is obvious<sup>(33)</sup>.

Many bands appeared at (1050-1400)  $\text{cm}^{-1}$  belong to the stretching vibration of  $\nu$  (C-N=N-C) also the band at (1000)  $\text{cm}^{-1}$  is for  $\nu$  (C-N)<sup>(34)</sup>.

The two mono substituted aromatic rings on the imidazole ring are indicated by the absorption at (690) and (760)  $\text{cm}^{-1}$ .

## Conclusions

- ❖ A new, simple, speed and sensitive method to determination of copper and ferric ions.
- ❖ Stability constants refer to the high stability of two complexes.
- ❖ These two complexes under optimum conditions are stable to more than 24 hour.

## Statistical Study

- ❖ Correlation factor  
Correlation factor between Cu and Fe was show in table (3)

**Table (3) Correlation factor between Cu and Fe**

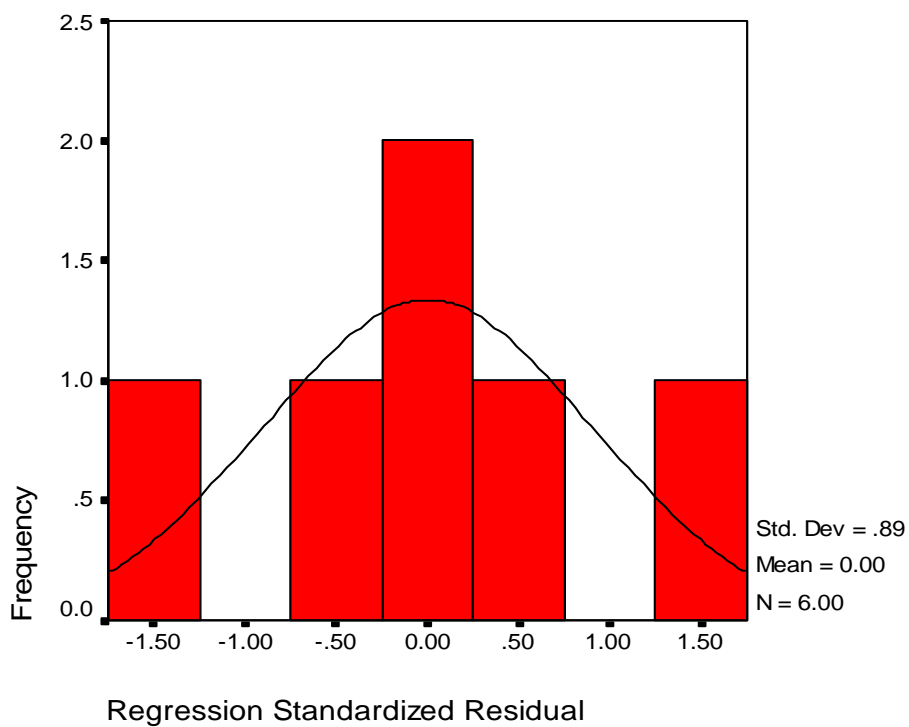
		Cu (II)	Fe (III)
Cu	Pearson Correlation	1	0.997**
	Sig. (2-tailed)	0	0.001
	N	6	6
Fe	Pearson Correlation	0.997**	1
	Sig. (2-tailed)	0	0.001
	N	6	6

\*\* Correlation is significant at the 0.01 level (2-tailed).

### ❖ Regression

The regression model of Cu was show in histogram (1) and figure (17)





Histogram (1) dependant variable (Cu)

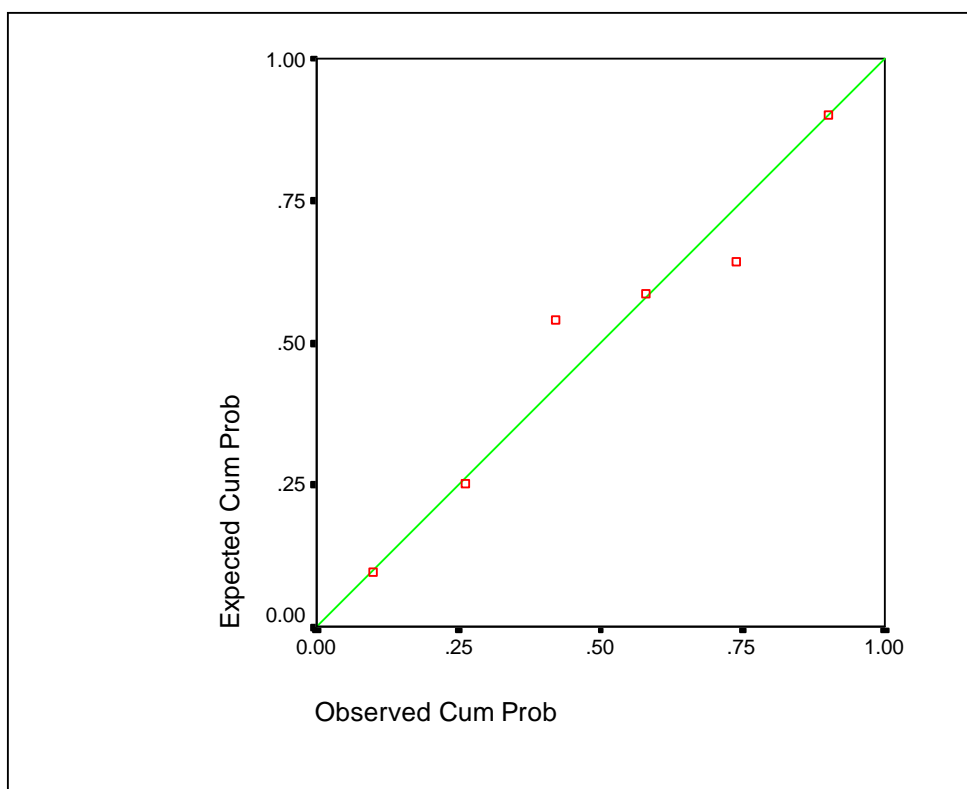
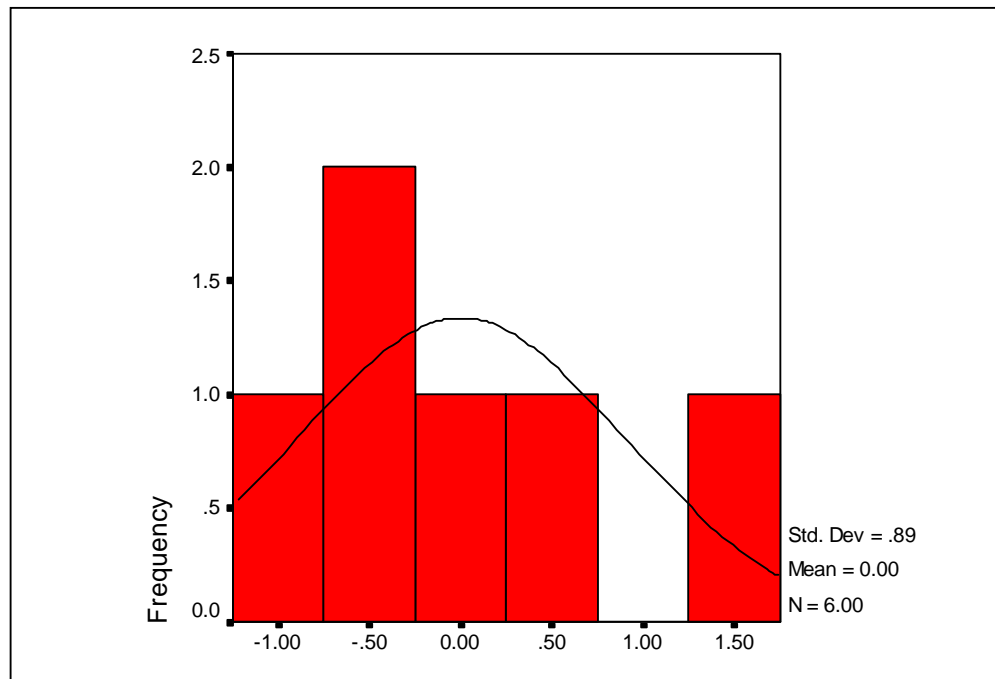


Figure (17) Normal P-P plot of regression standardized residual dependent variable (Cu)

The regression model of Fe was show in histogram (2) and figure (16)



Histogram (2) dependant variable (Fe)

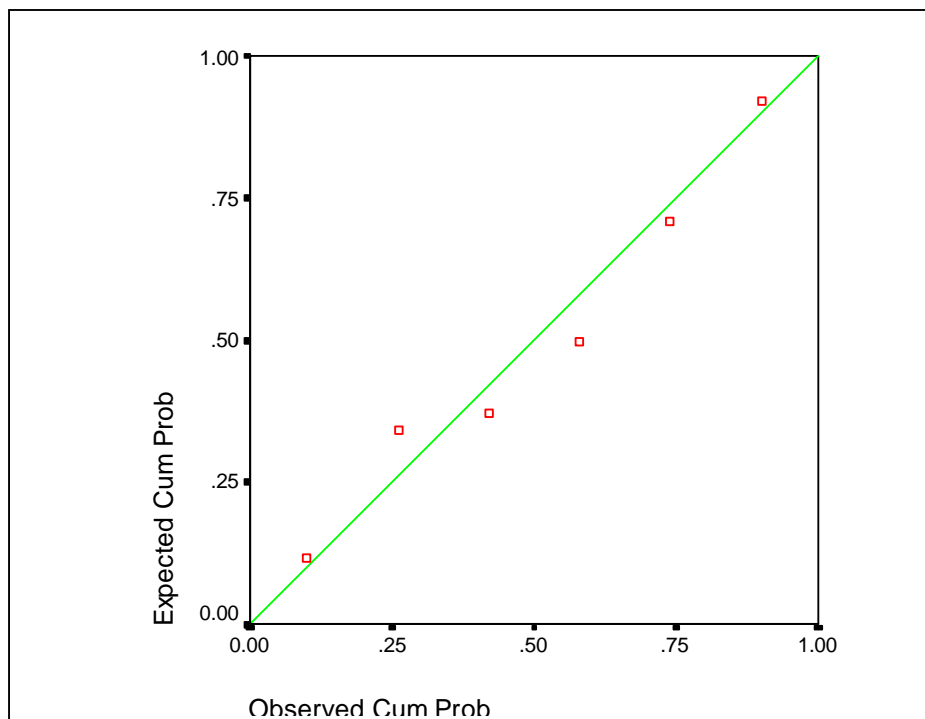


Figure (18) Normal P-P plot of regression standardized residual dependent variable (Fe)

## References

1. S. B. Savvin, V. P. Dedkova, O. P. Shvoeva, *Russ. Chem. Rev.* 2000, **69**, 187.
2. S. A. Morozko, V. M. Ivanov, *Zh.Anal.Khim.* 1996, **51**, 631.
3. S. A. Morozko, V. M. Ivanov, *Zh.Anal.Khim.* 1995, **50**, 629.
4. B. Kuswandi, A. A. Vaughan, R. Narayanaswamy, *Anal. Sci.* 2001, **17**, 181.
5. B. Kuswandi, R. Narayanaswamy, *Analytical Letters* 1999, **32**, 649.
6. M. L. Toral, N. Lara, J. Narvaes & P. Rich, *J.Chil. Chem. Soc.*, 2004, **49** (2), 163.
7. J. Savice, & V. Vasic, *Acta. Chem. Slov.*, 2006, **53**, 36.
8. L. Ferreira, A. queiroz, J. Assis, M. Korn & S. Gosta, *J. Braz. Chem. Soc.*, 1997, **8**(6), 621.
9. L. S. Jensen, *J Nutr* 1975, **105** (6), 769.
10. M. Thakur & M. Kdeb, *Talanta*; 1999, **49**, 561.
11. Y. Zhu, J. Liu, P. Liu & X. Zhou, *Mikrochem. Acta*, 1993, **112**, 127.
12. K. Ohshita, H. Wada & G. Nakagawa, *Anal.chem. Acta.*, 1985, **176**, 411.
13. A.A. Jewad, K.H. Kadhim & A.N. Al-Sharefy, *Nat. J. Chem.*, 2005, **19**, 331.
14. N. A. Fakhri, S.B. Dekran, H.A. Saleh & A.H. Alkadhumi, *J. of College of Edu., University of Salahaddin*, 1989, **1**, 19.
15. M. Zenki, Y. Lwadou, T. Yokoyhama, *Anal- Science*, 2002, **18**, 1077.
16. P. Richter, M.I. Toral, A.E. Tapia & E.Fuenzalida, *Talanta*, 1997, **122**, 1045.
17. F.W. Oehme, *Toxicity of Heavy Metals in the Environment*, Marcel Dekker, New York, 1978.
18. J.M. Moor, *Inorganic Contaminants of Surface Water*, Springer Verlag, New York, 1991.
19. S.B. Sonowale, Y.V. Ghalsasi & A.P. Argekar, *The Jap. Soc. Anal. Chem.*, 2001, **17**, 285.
20. M.N. Khan & A. Sarwar, *Anal. Science*, 2001, **17**(10), 1195.
21. M.J. Ahmed, I. Jahan & S. Bano, *Anal. Science*, 2002, **18**(7), 805.
22. J.F. Vanstaden & A. Botha, *Talanta*, 1999, **49**, 1099.
23. S. Nohut, S. Karaboecek, S. Guener & Y. Goek, *J. Pharm. Biomed. Anal.*, 1999, **20**, 309.
24. Y. Zhou & H. Jiang, *Fenxi Huaxue*, 1992, **20**, 1228.
25. S. Shibata, M. Furukawa & R. Nakaskima, *Analytical Chemical Acta*, 1976, **81**, 131.
26. P. Jop, *Ann. Chem*, (Paris), 1928, **9**, 113.
27. J.A. Yoe & A.L. Jones, *Chem. Anal. Ed.*, 1944, **11**, 16.
28. W.C. Vosburgh & G.R. Cooper, *J. Am. Chem. Soc.*, 1941, **63**, 437.
29. H. Shuker, *M.Sc. Thesis*, Mousl University, 1992.
30. C.B. Ojeda, A.G. Detorres, F.S. Rojas & J.M. Canopavon, *Analyst*, 1987, **112**, 1499.
31. W. Abd Al-Azezz, *Ph.D Thesis*, Mosul University, 1990.
32. F.N. Al-Obaidi, *Ph.D Thesis*, University College of North Wales, Bager, 1981.
33. P. Byabartta, S. Pal, T. Misra, C. Sinha, F. Liao, K.P. Selvan & T.H. Lu, *J. Coord. Chem.*, 2002, **55**, 479.
34. C. Preased & S. Benerji, *J. Inorg. Nucl. Chem.*, 1976, **83**, 1387.
35. D.M. White & J. Sonnenberg, *J. Org. Chem.*, 1924, **29**, 1942.