

Synthesis and Spectroscopic Characterization of New Coordination Compounds of Tin(IV) with Carbohydrazones

SUNITA CHOUDHARY, S. VARSHNEY and A. K. VARSHNEY*

Department of Chemistry, University of Rajasthan, Jaipur-302004, India

(E-mail: anilakv123@rediffmail.com)

ABSTRACT: Some new coordination compounds of tin(IV) having general formula $[\text{SnBu}(\text{HL})(\text{Cl})_3]$ (where HL = carbohydrazone ligand) have been synthesized by the reaction of monobutyltin(IV) trichloride with carbohydrazone ligands with the ratio of 1:1 (metal- ligand) using dry benzene as a reaction medium. The newly synthesized complexes were characterized by elemental analysis, molecular weight determinations and spectral analysis *viz.*, IR, UV-Vis and NMR (^1H , ^{13}C and ^{119}Sn). A distorted octahedral geometry of these complexes has been assigned on the basis of spectral studies. The biological activities of carbohydrazones and their Sn(IV) complexes have been screened *invitro* against some bacterial and fungal strain to assess their growth inhibitory potency. Most of the metal complexes exhibit more antibacterial and antifungal activities than the free carbohydrazone ligands against these organisms.

Keywords: Organotin(IV) complexes, carbohydrazones, spectral analysis, biological activities.

INTRODUCTION

Carbohydrazones are known to be a class of versatile ligands in coordination chemistry due to their ease of synthesis and diversity as well as structural possibilities. According to literature, they have been also as a potential antimicrobial, antioxidant, therapeutic, anticonvulsant, cytotoxic and pharmacological as well as catalytic agents.¹⁻⁴ Carbohydrazone ligands synthesizes using carbohydrazide having enormous

biological applications due to their oxygen and nitrogen donor atoms. Carbohydrazones exist in equilibrium of various tautomers due to unique structural features which greatly affect their chelating ability.⁵

The organometallic complexes of tin(IV) play a special role, due to their structural diversity. The chemistry of organotin(IV) derivatives is important due to their wide range of applications. The increasing importance of organotin(IV) complexes is due to their wide spread use in plastic industries such as PVC stabilizing agents⁶ and antineoplastic agents⁷ also broadly use in coating of ship hulls⁸ and marine antifouling agents.⁹ Organotin complexes are commonly used in agricultural field as wood preservatives¹⁰, fungicides¹¹ and insecticides¹². Organotin complexes exhibit pharmacological applications such as bacteriocides¹³ as an antitumor¹⁴⁻¹⁹ agents as an antimicrobial agents²⁰⁻²² as an anti-inflammatory agents²³ as an antiviral agents²⁴⁻²⁵ as an anti tuberculosis agents²⁶ and as an antihelmintics²⁷ and so on. Some of organotin(IV) derivatives containing O-O' bidentate donors showed strong antitumor activity than of cis-platin and mitomycin-C.²⁸

Continuing earlier research²⁹ on biologically active complexes, systematic studies on the binding of carbohydrazones to Sn(IV) metal ion lead to the conception to develop new and efficient complexes, play a vital role in a vast number of biological processes. In view of these facts, here we were thus motivated to undertake a systematic study of preparation and structural characterization of some new Sn(IV) complexes formed with carbohydrazones (HL) and Sn(IV) ion. In addition, the biological studies were applied to the free ligands and their Sn(IV) complexes against different bacterial, fungal strains using inhibitory zone diameter.

EXPERIMENTAL PART

Materials

Solvents were dried by standard methods³⁰ before use. *n*-BuSnCl₃ (Aldrich) has been used as supplied. The ligands used in this study have been synthesized in the laboratory using standard method reported in the literature.³¹

Physical measurements

Melting points were recorded using Gallenkamp melting point apparatus. Fourier transform infrared spectra (FTIR) were recorded using 8400 Shimadzu FT-IR Spectrophotometer using KBr pellets in the range of 4000-400 cm^{-1} . ^1H and ^{13}C NMR in CDCl_3 solution were recorded with a JEOL-FT A1 300 MHz spectrometer using TMS an internal reference. ^{119}Sn NMR spectra with proton noise decoupling in dry DMSO as the solvent, were recorded on a 90 MHz JEOL spectrometer using TMT (tetramethyltin) an internal reference. Micro elemental analysis (C, H, N) were recorded on a Coleman 5612 analyzer. The UV-Vis spectra of the carbohydrazones and their Sn(IV) complexes were recorded on a 1800 Shimadzu UV spectrophotometer in the range of 200-800 nm. Molecular weights were determined by Rast method. All the compounds have been synthesized using similar method therefore the synthetic and analytical data of the prepared complexes have been summarized in table 1.

Table 1. Physical data of Sn(IV) complexes of carbohydrazones

S. No	Empirical formula	% Yield	M.P. ($^{\circ}\text{C}$)	Color	% Elemental analysis				M:L in dry benzene	Mol. Wt Found (Calcd.)
					Found	(Calcd.)				
					C	H	N	Sn		
1	$\text{Sn}[(\text{C}_{20}\text{H}_{30}\text{N}_5\text{O}_2)(\text{Cl})_3]$	65	135	Brown	40.20 (40.23)	5.06 (5.06)	11.73 (11.72)	9.20 (9.19)	1:1	597.02 (597.53)
2	$\text{Sn}[(\text{C}_{23}\text{H}_{30}\text{N}_5\text{O}_2)(\text{Cl})_3]$	70	141	Gray	43.62 (43.60)	4.77 (4.77)	11.06 (11.05)	8.67 (8.67)	1:1	633.32 (633.56)
3	$\text{Sn}[(\text{C}_{18}\text{H}_{26}\text{N}_5\text{O}_2)(\text{Cl})_3]$	63	150	Dark green	37.98 (37.96)	4.60 (4.60)	12.30 (12.30)	9.65 (9.65)	1:1	569.23 (569.48)
4	$\text{Sn}[(\text{C}_{22}\text{H}_{28}\text{N}_5\text{O}_2)(\text{Cl})_3]$	74	147	Dark brown	42.67 (42.65)	4.56 (4.55)	11.31 (11.30)	8.87 (8.87)	1:1	619.17 (619.54)
5	$\text{Sn}[(\text{C}_{19}\text{H}_{28}\text{N}_5\text{O}_2)(\text{Cl})_3]$	68	130	Green	39.12 (39.11)	4.84 (4.83)	12.00 (12.00)	9.42 (9.41)	1:1	583.24 (583.51)

Synthesis of carbohydrazones

Carbohydrazones have been synthesized by refluxing of the reaction mixture using a hot ethanolic solution of carbohydrazide (2.5 mmol) and hot ethanolic solution of suitable carbonyls (2.5 mmol) *viz.*, salicylaldehyde, 2-hydroxy-1-naphthaldehyde, *o*-hydroxyacetophenone, 2-hydroxy-1-acetonaphthone, *o*-

hydroxypropiophenone and 2-acetylpyrrole in 1:1:1 M ratio for 2 hrs. The desired products were obtained after evaporation of the solvent were filtered and then recrystallized from ethanol.

Synthesis of complexes A¹-A⁵ of carbohydrazones

Reaction of monobutyltin(IV) trichloride (0.500 g, 1.78 mmol) with ligand (0.555 g, 1.78 mmol) have been carried out in 1:1 M ratio in dry benzene. The reaction mixture was heated under reflux on a fractioning column for 8-10 hrs. After the completion of reaction, the excess solvent was removed under reduced pressure to yield colored viscous liquid (yield 63-74 %). It was purified by *n*-hexane/benzene mixture.

Antimicrobial activity

Antimicrobial activity of chemically derived compounds was studied. Three bacterial and fungal strains were selected for the primary screening.

Microorganisms Used

Clinical laboratory bacterial isolates of *Bacillus subtilis*, *Staphylococcus aureus*, and *Escherichia coli* and fungal isolates viz., *Fusarium oxysporium*, *Trichoderma reesei* and *Penicillium funiculosum* were collected from the stock cultures of Microbiology Laboratory, SMS Medical College Jaipur, India.

Preparation of samples

The 10 mg/mL of samples was dissolved in DMSO and further dilutions were made for calculating MIC value.

Culture and maintenance of bacteria

Pure cultures obtained from SMS Medical College Jaipur, India were used as indicator organisms. These bacteria were grown in Nutrient agar medium (prepared by autoclaving 8 % Nutrient agar of Difco-Laboratories, Detroit, USA, in distilled water at 15 lbs psi for 30 min) by incubating at 37 °C for 48 hrs. Each bacterial culture was further maintained on the same medium after every 48 hrs of transferring. A fresh suspension of test organism in saline solution was prepared from a freshly grown agar slant before antimicrobial assay.

Determination of Antibacterial Assay

In vitro antibacterial activity of the samples was studied against gram positive and gram negative bacterial strains by the agar well diffusion method.³² Streptomycin was used as reference antibacterial agent (control). Mueller Hinton agar no. 2 (Hi Media, India) was used as the bacteriological medium. The Mueller Hinton agar was melted and cooled to 48–50 °C and a standardized inoculum (1.5×10^8 CFU/mL, 0.5 McFarland) was then added aseptically to the molten agar and poured into sterile petri dishes to obtain a solid medium. Wells were prepared in the seeded agar plates. The test compound (100 μ L) was introduced in the well (6 mm). The plates were incubated overnight at 37 °C. The antimicrobial spectrum of the chemical compounds was determined for the bacterial species in terms of zone sizes around each well. The diameters of zone of inhibition produced by the agent were compared with those produced by the commercial control antibiotics, streptomycin. For each bacterial strain controls were maintained where pure solvents were used instead of the chemical compound. The control zones were subtracted from the test zones and the resulting zone diameter was measured with antibiotic zone reader to nearest mm. The experiment was performed three times to minimize the error and the mean values are presented.

Determination of Antifungal Assay

Anti-fungal activity of the experimental plant was investigated by agar well diffusion method.³³ Ketokenazole was used as reference antifungal agent. The yeasts and saprophytic fungi were sub cultured onto Sabouraud's dextrose agar, SDA (Merck, Germany) and respectively incubated at 37 °C for 24 hrs and 25 °C for 2-5 days. Suspensions of fungal spores were prepared in sterile PBS and adjusted to a concentration of 10^6 cells/mL. The plates were dried at room temperature for 15min. Wells of 10 mm in diameter and about 7 mm apart were punctured in the culture media using sterile glass tube. 0.1 ml of several dilutions of fresh extracts was administered to fullness for each well. Plates were incubated at 37 °C. After incubation of 24 hrs bioactivities were determined by measuring the diameter of inhibition zone (in mm). All experiments were made in triplicate and means were calculated.

RESULTS AND DISCUSSION

The condensation reaction of carbohydrazide with suitable carbonyls in 1:1:1 M ratio yields carbohydrazones.

Sn(IV) complexes of carbohydrazones

These have been synthesized via reaction of *n*-butyltin(IV) trichloride with carbohydrazones in 1:1 M ratio, in refluxing with dry benzene which yielded corresponding coordination compounds of Sn(IV). All these synthesized compounds are colored viscous liquid and monomeric in nature. The analytical data of carbohydrazones and their metal complexes are given at table (1), in a satisfactory agreement with the calculated values.

UV-Vis spectra

The UV-Vis spectra of the free ligands and their complexes were recorded using ethanol at room temperature. The UV spectrum of carbohydrazones (HL) showed two intense bands at 225-290nm and 370-410 nm which belong to $\pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ transition respectively (Table 2). In spectra of Sn(IV) complexes, the band at 225-290 nm can be attributed to $\pi \rightarrow \pi^*$ transitions, which were shifted towards lower energy region (245-295 nm). This was a result of coordination of carbohydrazone ligand to the tin center. The band around 370-410 nm can be assigned for free ligands and it was shifted to 345-385 nm in complexes due to coordination of azomethine nitrogen to the tin atom.³⁴

Table 2. UV-Vis Spectra of (HLs) and their metal complexes.

Ligands	UV-Vis bands (nm)	Complexes	UV-Vis bands (nm)
HL ¹	225, 370	A ¹	245, 345
HL ²	234, 382	A ²	275, 357
HL ³	250, 387	A ³	295, 365
HL ⁴	265, 399	A ⁴	265, 373
HL ⁵	290,410	A ⁵	270, 385

Infrared Spectroscopic Study

Reaction of carbohydrazide with suitable carbonyls produces carbohydrazones (HL). This reaction was followed by appearance of a characteristic new band in the range 1605-1625 cm^{-1} , which is due to frequency of the free azomethine groups, $\nu_{\text{C=N}}$ are utilized to confirm the structures of (HL).³⁵ The bands due to $\nu_{\text{C=N}}$, were shifted to lower frequencies showing the coordination of nitrogen to tin ion.³⁶ A broad peak due to ν_{OH} of -OH group appear in the region 3405-3450 cm^{-1} in the spectra of ligands. In spectra of Sn(IV) complexes this band is shifted at 3320-3370 cm^{-1} . This indicates the involvement of oxygen of this group in bonding with tin metal. The medium to sharp intensity bands are observed around 520-550 cm^{-1} and 585-600 cm^{-1} , which may be assigned to the asymmetric and symmetric mode of $\nu_{\text{Sn-C}}$ stretching vibrations in the spectra of tin complexes and also two bands at 525-555 cm^{-1} and 410-430 cm^{-1} may be assigned to $\nu_{\text{Sn-O}}$ and $\nu_{\text{Sn}\leftarrow\text{N}}$ vibrations, respectively, indicating the participation of azomethine and phenolic oxygen in complexation. The $\nu_{\text{C-O}}$ and $\nu_{\text{(N-N)}}$ medium to strong intensity bands appear at 1260-1270 cm^{-1} and 980-990 cm^{-1} respectively, these bands are also slightly shifted to higher frequency region as a result of complex formation, showing chelation of oxygen atom to the tin atom.³⁷ IR spectral data are systemized at table 3.

Table 3. Characteristic stretching vibration frequencies (cm^{-1}) located at FT-IR of (HLs) and their metal complexes.

Comp. No.	$\nu_{\text{C=N}}$	$\nu_{\text{Sn-N}}$	$\nu_{\text{Sn-O}}$	$\nu_{\text{N-N}}$	ν_{OH}
HL ¹	1605	-	-	980	3405-3450
HL ²	1610	-	-	983	3410-3440
HL ³	1622	-	-	987	3415-3435
HL ⁴	1625	-	-	990	3408-3430
HL ⁵	1615	-	-	985	3405-3445
A ¹	1575	415	550	995	3320-3350
A ²	1582	412	545	997	3315-3340
A ³	1578	425	555	1005	3330-3370
A ⁴	1590	430	530	1002	3310-3345
A ⁵	1595	410	525	999	3335-3360

¹H NMR

The ^1H NMR data of the used ligands and their organotin(IV) complexes were recorded in CDCl_3 (Table 4). For ^1H NMR spectra of ligands, the signal due to the OH proton of the ligands appears at δ 11.10-12.05 ppm (S), was shifted downfield in the spectra of the corresponding tin complexes showing thereby chelation of the ligand moiety through the phenolic oxygen to the tin atom (δ 11.25-12.35 ppm). In the case of the ligands, the proton signal for the methyl protons $[-\text{C}(\text{CH}_3)=\text{N}]$ and azomethine protons $[-\text{CH}=\text{N}]$ in the region δ 1.78-1.85 ppm (S) and δ 8.10-8.30 ppm, shifts downfield in the spectra of corresponding tin complexes on account of its deshielding, which is attributed to the donation of the lone pair of electrons by the azomethine nitrogen to the tin atom. The ligands show a complex multiple in the region δ 6.09-7.45 ppm for the aromatic protons which remains at almost the same position in the same spectra of the organotin(IV) complexes. However, a broad signal at δ 3.30-3.77 ppm for the NH_2 protons remains almost unaltered in the tin complexes which clearly show that this group does not take part in the complexation reaction. The complexes, however, show additional signals at δ 0.75-1.95 ppm owing to the protons of the butyl group.

Table 4. ^1H NMR data for the ligands and for their corresponding organotin(IV) complexes

Comp. No.	-HCN	-OH	$-\text{CH}_3\text{CN}$	$-\text{NH}_2$	Aromatic protons
HL ¹	8.10	11.10	1.78	3.30	6.18-7.21
HL ²	8.15	11.50	1.81	3.53	6.09-7.14
HL ³	8.30	11.80	1.85	3.77	6.67-7.34
HL ⁴	8.20	12.02	1.83	3.48	6.10-7.45
HL ⁵	8.25	12.05	1.79	3.40	6.78-7.38
A ¹	8.60	11.25	1.90	3.35	7.23-7.78
A ²	9.10	11.65	1.93	3.60	7.15-7.70
A ³	9.25	12.20	2.01	3.80	7.68-7.95
A ⁴	9.00	12.28	1.99	3.56	7.34-7.89
A ⁵	9.08	12.35	1.97	3.48	7.46-7.87

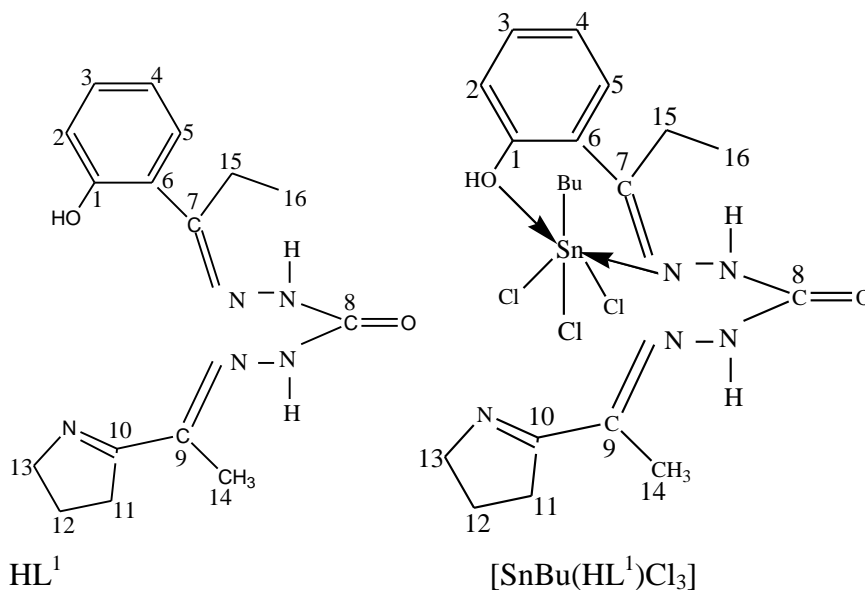
^{13}C NMR Spectra

The ^{13}C NMR spectral data for carbohydrazones and their corresponding tin complexes are reported in table 5. The shifting in position of resonance of carbon

attached to OH group suggests the bonding of oxygen to the tin atom. The signal due to the carbon atom attached to the azomethine group in the ligands appears at δ 172.6-174.7 ppm. Further the shifting of azomethine ($>C=N$) carbon signal in the spectra of complexes (δ 165.1-166.8 ppm) indicate that the azomethine nitrogen has been involved in coordination with the tin atom. The carbon of butyl group is observed at δ 14.5-29.2 ppm which is comparable to other similar tin compounds.

Table 5. ^{13}C NMR data for the ligand and for its corresponding organotin(IV) complexes

Comp.	Chemical shift value in δ ppm															
	C-1	C-2	C-3	C-4	C-5	C-6	C-7	C-8	C-9	C-10	C-11	C-12	C-13	C-14	C-15	C-16
HL ¹	155.5	116.1	128.1	118.5	127.2	117.3	174.3	168.3	173.8	142.2	11.2	11.7	12.1	12.3	12.6	11.9
A ¹	165.3	115.4	129.7	119.9	128.3	119.4	165.9	184.8	165.3	143.4	10.9	11.1	11.9	13.8	13.3	12.1



^{119}Sn NMR Spectra

^{119}Sn NMR spectra of tin complexes have been recorded using TMT (tetramethyltin) as external standard. Tin (IV) complexes give sharp signals at δ -350.6 ppm, which is in accordance with the proposed six coordinated distorted octahedral geometry, in agreement with the previously reported values.³⁸ Chemical shift values for similar six coordinated BSn(IV) complexes have been reported in the range of δ -270.6 to -370.4 ppm.

In-Vitro antimicrobial assay

The *in-vitro* antibacterial activity of the ligands and their complexes were screened against three bacterial strains by well diffusion method using streptomycin as reference they showed significant potent activity. The susceptibility of bacterial strain toward the compounds was estimated by measuring the size of inhibition zone diameter. It is evident from the results that the antibacterial activity of some of the metal complexes is higher than the free ligands and lesser than the standard against all the bacteria tested. In case of antibacterial activity carbohydrazones and their metal complexes were found to be active. Antifungal activity of the ligands and their complexes were screened against three fungal strains by well diffusion method using Ketokenazole as reference they showed some potent activity. The antimicrobial results are systemized at table 6. It is, however, known that the chelating tends to make carbohydrazones act as more powerful and potent bacterostatic agents, thus, inhibiting the growth of bacteria and fungi more than the parent carbohydrazones.

Table 6. Antimicrobial results of carbohydrazones and their metal complexes.

Ligands/ Compounds	Antibacterial activity zone of inhibition (mm)					Antifungal activity zone of inhibition (mm)				
	Bacterial strain	20 mL	40 mL	60 mL	80 mL	Fungal strain	20 mL	40 mL	60 mL	80 mL
HL ¹	<i>B. subtilis</i>	9	11	15	18	<i>F. oxysporium</i>	Nill	nill	11	14
	<i>S. aureus</i>	10	13	14	16	<i>T. reesei</i>	nill	10	14	18
	<i>E. coli</i>	9	14	15	18	<i>P. funiculosum</i>	8	10	13	16
HL ²	<i>B. subtilis</i>	9	12	15	17	<i>F. oxysporium</i>	9	10	12	14
	<i>S. aureus</i>	10	14	16	18	<i>T. reesei</i>	8	10	14	16
	<i>E. coli</i>	9	11	14	16	<i>P. funiculosum</i>	9	12	14	16
HL ³	<i>B. subtilis</i>	8	11	14	15	<i>F. oxysporium</i>	nill	10	12	17
	<i>S. aureus</i>	10	13	14	17	<i>T. reesei</i>	8	9	10	13
	<i>E. coli</i>	9	12	15	16	<i>P. funiculosum</i>	9	12	13	16
HL ⁴	<i>B. subtilis</i>	9	10	12	14	<i>F. oxysporium</i>	8	11	12	13
	<i>S. aureus</i>	10	12	13	15	<i>T. reesei</i>	8	12	11	14
	<i>E. coli</i>	11	12	13	16	<i>P. funiculosum</i>	9	10	12	14
HL ⁵	<i>B. subtilis</i>	9	10	13	15	<i>F. oxysporium</i>	9	10	13	15
	<i>S. aureus</i>	10	13	14	16	<i>T. reesei</i>	8	9	11	14
	<i>E. coli</i>	11	12	14	15	<i>P. funiculosum</i>	nill	nill	12	16

A ¹	<i>B. subtilis</i>	12	13	13	16	<i>F. oxysporium</i>	10	12	14	nill
	<i>S. aureus</i>	12	14	15	20	<i>T. reesei</i>	nill	nill	8	13
	<i>E. coli</i>	10	12	14	16	<i>P. funiculosum</i>	10	12	14	16
A ²	<i>B. subtilis</i>	10	13	15	18	<i>F. oxysporium</i>	10	11	14	16
	<i>S. aureus</i>	12	14	17	18	<i>T. reesei</i>	10	12	13	14
	<i>E. coli</i>	10	12	16	17	<i>P. funiculosum</i>	12	14	14	16
A ³	<i>B. subtilis</i>	9	11	14	16	<i>F. oxysporium</i>	8	12	13	16
	<i>S. aureus</i>	11	12	13	18	<i>T. reesei</i>	10	10	12	14
	<i>E. coli</i>	12	14	16	17	<i>P. funiculosum</i>	10	10	15	18
A ⁴	<i>B. subtilis</i>	10	11	13	15	<i>F. oxysporium</i>	10	12	14	16
	<i>S. aureus</i>	11	12	13	15	<i>T. reesei</i>	12	14	16	18
	<i>E. coli</i>	11	13	14	17	<i>P. funiculosum</i>	10	12	14	16
A ⁵	<i>B. subtilis</i>	10	12	15	19	<i>F. oxysporium</i>	9	11	13	17
	<i>S. aureus</i>	10	12	16	17	<i>T. reesei</i>	10	10	14	16
	<i>E. coli</i>	12	13	18	16	<i>P. funiculosum</i>	9	10	13	14

Streptomycin (for bacteria) – Inhibition zone – 20mm; Ketokenazole (for fungi) – Inhibition zone – 22mm. *Standard

Structure of Sn(IV) complexes A¹-A⁵

On the basis of above spectral analysis, a shift in the position of O-H, C-O, N-N and –C=N groups suggest the bidentate nature of ligands. In all Sn(IV) complexes (Fig. 1), in view of the presence of one bidentate chelate rings, three chlorine atoms, one butyl group and monomeric nature of these complexes, the following structure is being proposed in which central tin atom acquires distorted octahedral geometry.

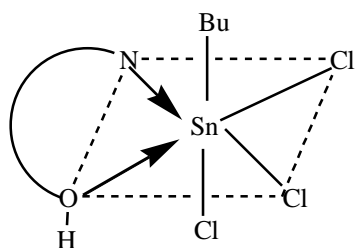


Figure- 1: Sn(IV) complexes ([A₁] – [A₅])

CONCLUSIONS

In summary, we have synthesized some new adducts of monobutyltin(IV) with carbohydrazone ligands with determination of their structures and physical properties. FTIR and NMR spectral studies suggest the bidentate nature of ligands, which coordinate with Sn(IV) ions through azomethine nitrogen and phenolic oxygen.

The distorted octahedral geometry of these compounds of the type [SnBu(HL)(Cl)₃] (where L = carbohydrazone ligand) have been proposed on the basis of elemental analysis, spectral studies *viz.* IR, UV-Vis and NMR (¹H, ¹³C and ¹¹⁹Sn). Furthermore, these Sn(IV) complexes were found to have significant antibacterial and antifungal activity.

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